

ANTIDEPRESSANT AND PRO-REPRODUCTIVE EFFECTS OF *MENTHA SPICATA* & *TERMINALIA CATAPPA* EXTRACTS IN STEROID-RESISTANT PCOS – A CASE REPORT AND LITERATURE REVIEW

<sup>1,\*</sup>Ajibola Monisola UMARUDEEN, <sup>2</sup>Chinonye Anuli MADUAGWUNA and <sup>3</sup>Oluchi Ngozika OKOLI

<sup>1,2</sup>Department of Pharmacology & Therapeutics, Faculty of Basic Clinical Sciences, University of Abuja, Nigeria

<sup>3</sup>Department of Obstetrics & Gynaecology, Faculty of Clinical Sciences, University of Abuja, Nigeria

Received 07<sup>th</sup> January 2026; Accepted 10<sup>th</sup> February 2026; Published online 30<sup>th</sup> March 2026

Abstract

**Introduction:** Polycystic ovary/ovarian syndrome (PCOS) is the commonest reproductive endocrinopathy in women. Amenorrhea/oligomenorrhea, hirsutism, and bilateral polycystic ovaries are its dominant symptoms. **Materials & Methods:** This study is both a consented report on the history, clinical, biochemical and radiological features of on a 25-year woman who presented to us and a literature review on PCOS. Her presenting features included moderate anaemia, amenorrhea/oligomenorrhea, dysmenorrhea, hirsutism, and bilateral polycystic ovaries. Other included prolonged/heavy menorrhagia which started after being placed on monthly oral contraceptive pills from her previous medical consultations before presenting to us. She volunteered that the latter features were often associated with mood swings and anxiety. Her overall profile indicated a diagnosis of PCOS with moderate depressive symptoms. The contraceptive pills were tailed off while she was immediately given haematinics and nutritional support. She was subsequently placed on a twice daily hot water decoctions of 2.5 g *Mentha spica* (MS) and teaspoonful of *Terminalia catappa* (TC) powders for several weeks/months. About 7 days of commencing the herbal treatment, clinical improvement was observed in menorrhagia, and 8 days later, the vaginal bleeding ceased. Although her hirsutism and dysmenorrhea did not improve very significantly, her moods, anxiety traits, menstrual cycles and flows progressively regularized until she achieved cyesis 9 months into the herbal treatment. In the meantime, Internet was also searched using several key words centered on PCOS treatments, aetiopathogenesis, and epidemiology. Full articles were selected, perused, and pcos-relevant information was included in the report. **Findings:** This study shows PCOS is a highly prevalent multi-endocrine, multi-genetic, and multifactorial gynaecological disorder with no streamlined standard drug treatments. The study further indicates the oral hormone contraceptive pills – the first/default and other synthetic anti-pcos drugs have efficacy and toxicity liabilities. This study suggests MS and TC extracts may have anti-pcos effect. **Conclusion:** This may justify the traditional use of extracts of these and other medicinal plants for the treatment of PCOS.

**Keywords:** PCOS, Spear mint, Tropical almond, Menorrhagia, Powder, Decoction

INTRODUCTION

Polycystic ovary syndrome (pcos) or Stein-Leventhal syndrome is a disorder having oligo/amenorrhea, hirsutism, and multiple cysts in both ovaries as its most prominent clinical features. From the original description by Irving Freiler Stein and Michael Leventhal in 1935, the syndrome is now recognized as a complex multi-endocrine disorder characterized by features of hyperandrogenism, insulin insensitivity, dysfunctional hypothalamus-pituitary-adrenal axis, and hypo-functioning multiple cysts-containing (polycystic) ovaries. Bilateral polycystic ovary morphology results from accumulated unruptured immature Graffian follicles, a fallout from recurrent anovulatory menstrual cycles (1–4)

The syndrome is currently regarded as the commonest gynaecology-related endocrinopathy in reproductive age. Currently, an estimated 10% or 200 million of the global women population reportedly suffer from pcos. The most frequent clinical features of this syndrome to date still include irregular, infrequent, anovulatory, or absent menstrual cycles (ovarian hypogonadism features), acne, male-type hirsutism and alopecia (hyperandrogenism features), dyslipidemia, hyperglycemia, and obesity (metabolic features) (5–7).

METHODOLOGY

For the literature review component of this report, data bases such as Google, Google scholars, and Pubmed were searched for literature relevant to pcos and anti-pcos treatments, drugs, phytochemicals, medicinal plants and non-drug modalities validated for the management of pcos. The key search words for surfing the internet included but not limited to “polycystic ovary syndrome”, “polycystic ovarian syndrome”, “polycystic ovary disorder”, “polycystic ovarian disorder”, “PCOS”, “PCOD”, “medicinal plants for PCOS”, “treatment of PCOS”, “drug treatment of PCOD”, “treatment of PCOD”, “drug treatment of PCOS” “medicinal plants for PCOD”, “phytochemicals used for PCOS”, “phytochemicals used for PCOD”, “herbs for PCOS”, “herbs for PCOD”, “herbal treatments for PCOS”, “herbal treatments for PCOD”, “*Mentha spicata* pcos”, “spearmint pcos”, “*Terminalia catappa* pcos”, “tropical almond”, “pcos treatments”, “treatment of PCOD”, “prevention of PCOS”, and “prevention of PCOD”. Journal/conference articles were searched for data/texts relevant to pcos, pcos treatments, *Mentha spicata*, spearmint, *Terminalia catappa*, tropical almond, were selected into the final write-up. For the Case Report component, personal, family, and past medical/drug treatment history as obtained from the client, clinical and laboratory examination findings, diagnosis and treatments as well as personal and clinical observations during treatments were all captured into the write-up. Relevant ethical clearance and full consent of the

\*Corresponding Author: Ajibola Monisola UMARUDEEN, Department of Pharmacology & Therapeutics, Faculty of Basic Clinical Sciences, University of Abuja, Nigeria.

client to the diagnosis, treatments, and this publication were duly obtained.

## PCOS AETIOPATHOGENESIS

Although both external (epigenetics, environmental toxicants, diets, stress, etc.) and internal (oxidative stress, insulin resistance, inflammation, obesity, and genetic) factors are frequently linked to the emergence of this disorder, so far, its complete aetiopathogenesis is not yet known (6–8). However, hyperandrogenism appears to be central to the pathogenesis of this disorder and to be at the convergence of most of the above-listed pcos contributory factors – particularly hyperinsulinemia and inflammation. Other factors not included in the list of pcos factors above, like high body gonadotropin-releasing hormone (GnRH) level, and high luteinizing hormone (LH) to follicle-stimulating hormone (FSH) ratio promote pcos emergence via hyperandrogenism resulting from increased steroidogenesis (6–8).

Scientific reports indicate pcosis an inherited polygenic trait whose phenotypic expression essentially requires exposure to any of the requisite afore-listed environmental risk factors – with genetics contribution to its inheritability currently estimated to be only around 10 percent(6,9–12). Genome-wide association studies (GWAS) imply over-expression of or polymorphisms in the promoter genes associated with the coding of certain proteins i.e., sex hormone binding globulin (SHBG), Thyroid Adenoma-Associated Protein (THADA); Variable Number Tandem Repeats Insulin ((VNTR); Fat Mass/Obesity (FTO); Insulin Receptor (INSR); Luteinizing Hormone/Chorionic Gonadotropin Receptor (LHCGR); DENN Domain-containing Protein 1 (DENND1A); and Peroxisome Proliferator-Activated Receptor (PPAR) are some of the genetic predispositions to pcos pathogenesis via enhanced ovarian androgen steroidogenesis, insulin dysregulation, lipogenesis, and inflammation (13–15). Research has also shown defects in the mitochondrial transfer RNA and increased genetic polymorphisms in pcos victims are associated with altered electron transfer and exaggerated reactive oxygen species generation, inflammation, and insulin resistance, compared to their non-pcos counterparts, Additional evidence suggests the 100% mitochondrial DNA heritability exclusively from mothers may be the genetic reason behind why pcos tends to run in families and siblings tend to share similar metabolic, androgenic, and menstrual features(15–17).

Epigenetics, non-DNA sequence-based modifications in genetic expression, is also thought to play a significant role in pcos pathophysiology. Both animal and human studies indicate that in-utero exposure of fetuses to high maternal levels of insulin resistance, insulin, androgens, and anti-mullerian hormone (AMH), independent of diets, is significantly linked to the emergence of pcos in adult life (16,17). Research indicates these fetal environmental factors are associated with increased fetal androgenesis resulting from hypomethylation and the resultant over-expression of fetal genes concerned with androgen and insulin metabolic up-regulation on one hand, and with reduced placental aromatase activity-linked testosterone breakdown, on the other (17–19). These effects collectively result in increased fetal androgen steroidogenesis. Another important epigenetic contribution to this disorder is viewed to come from the hypomethylation of certain genes involved in mitochondrial metabolism i.e., Parkinson disease associated (PARK2), estradiol/estrogen receptor1 (ESR1), and insulin

(INS) genes, all of which are partly involved in mitochondrial metabolism. The altered methylation of the latter genes has been associated with exaggerated rates of mitochondrial injury and death, accumulation of reactive species, lipogenesis, and apoptosis activation (20,21). Additionally, epigenetic re-programming of fetal DNA for adult life pcos emergence is suspected to come from the fetal DNA modulation by some micro RNAs. DNA modulatory activities of these micro RNAs (mi-155, mi-146a, mi-122, mi-132, mi-93, mi103, etc.,) are thought to be linked to pcos pathologic processes i.e., altered inflammation, insulin, androgen and lipid metabolism and regulation (22, 23).

Studies have shown chronic low-grade body systems-wide inflammation and cellular oxidative stress are ultimately pro-androgenic. Women with pcos are reported to exhibit greater rates of mitochondrial structural and metabolic anomalies with attendant greater risk of cellular oxidative stress, and reactive oxygen and nitrogen species generation from environmental stressors (e.g., psychological stress, infectious agents, apoptotic and senescent cells, chemicals, excessive energy intake, and accumulation of metabolic intermediates)(23–26). The increased rates of free reactive species are viewed to trigger diverse immune reactions that result in chronic systemic inflammatory processes (27,28). Systemic inflammation in pcos is also thought to arise from the activities of inflammasomes and other inflammatory propagators – cyclophilin A, adipokines, nuclear factor kappa B (NF- $\kappa$ B), and vascular dysfunction mediators (27,29–32). This is in addition to the evidence showing chronic systemic inflammation resulting from cellular oxidative stress, alone, or in concert with insulin resistance/insulinemia promotes hyperandrogenism in pcos women by upscaling ovarian steroidogenic enzymatic activities (33–35).

Research further shows although Luteinising hormone (LH) and insulin exhibit some synergistic actions in the generation of the hyperandrogenism seen in pcos, nevertheless each hormone can contribute to this independently (36–39). For instance, Insulin contributes to pcos hyperandrogenism in several ways – including direct mechanisms by the promotion of chronic systemic low-grade inflammation and stimulation of PI3K and MAPK pathways-linked ovarian androgenesis. Insulin also exerts indirect androgenic effects via increased GnRH-LH and hypothalamic-pituitary-adrenal (HPA) axis activity with the resultant increased ovarian steroidogenesis and insulinemia-enhanced inhibition of SHBG hepatic synthesis – thus increasing free (biologically active) plasma testosterone(37–42).

On LH part, it contributes directly to pcos hyperandrogenism by stimulating increased production of ovarian and adrenal ketosteroid precursors by Cytochrome P450c17 (CYP17A1) enzyme – whose expression is regulated by LH – and for this effect, LH's role in pcos hyperandrogenism was thought to be primary (4,43,44). However, many of LH's indirect hyperandrogenic effects arising from exaggerated LH serum levels, pulse frequency and amplitude often occur only in about half of all pcos cases. Thus, LH dysregulation, just like insulin resistance, is no longer considered the primary pathology responsible for hyperandrogenism in this syndrome. However, evidence shows that in the face of co-existing hyperinsulinism, the normal ovarian theca cell desensitization to LH stimulation at submaximal serum androgen levels is lost and the ovarian tissues are often hypersensitive to the

increased steroidogenic stimulation by LH's exaggerated levels (45–47). Nutrition is another factor implicated in the pcos process. This is because human nutrition involves a lifelong intake of diverse chemical substances, many of which are pro-/anti-inflammatory and/or pro-/antioxidant in nature. Diets are therefore widely considered an influential factor to the emergence/progression of pcos – a disease process known to be significantly hinged on cellular inflammation and oxidation. Thus, chronic consumption of alcohol (with acetaldehyde as a byproduct), small molecule sugars (glucose, fructose), advanced glycation end-products (AGEs), and short-chain fatty acids have been implicated in pcos aetiopathogenesis (9,10,12,13) because of a pro-androgenic pressure they exert via their negative impact on the gut microbiota as well as on SHBG synthesis and plasma levels (48–50). Current evidence implies chronic dietary exposure to these substances results in a pro-inflammatory shift in the host immunity, gut microbiota and wall integrity – ultimately leading to the generation of a chronic low-grade systemic inflammation and increased androgenesis and the inhibition of the hepatocyte nuclear factor-4 $\alpha$  (4HNF-4 $\alpha$ )-mediated SHBG synthesis, which in turn results in increased plasma levels of biologically active androgens (10,12,47,51–54).

Dysregulation of an important enzymatic pathway - the CYP enzyme complex - involved in ovarian/adrenal steroidogenesis is a major source of hyperandrogenism in pcos. This genetically determined functional dysregulation implicates both cytochrome P450 CYP17 and cytochrome P450 CYP19 involved in the conversion of androgens to sex ketosteroid precursors and formation of estrogen from androgens, respectively. Both conversions which are the rate-limiting steps in ovarian and adrenal steroidogenesis are known to have their enzymatic activities upregulated in pcos (53–55). Specifically, scientific reports indicate the increased androgen biosynthesis by 17, 20-lyase of CYP17 is due to the following factors: firstly, abundant presence of hemoprotein cytochrome P450 oxidoreductase (POR), secondly, greater cytochrome b5-facilitated POR-17, 20-lyase CYP17 allosteric interactions, and lastly, greater phosphorylation of 17, 20-lyase CYP17 in ovarian tissues (56–58). Dysregulation of the androgen-oestrogen catalytic activity of the CYP19 (ovarian aromatase) in pcos is associated with hypo-oestrogenism, hyperandrogenism, and disruptions of several metabolic processes for which it is essential (59–61). Research also implies that most if not all pcos biochemical (hyperandrogenaemia, oestrogenaemia), histological/radiological (impaired folliculosis, anovulation, pcos ovarian morphology), and clinical (hirsutism, male pattern of abdominal/chest hair distribution) features are reproducible with hyperandrogenism induced by aromatase dysfunction – suggesting ovarian aromatase dysregulation may be essential to pcos aetiopathogenesis (61–63).

Classification of pcos is as complex and un-unified as the clinical presentations of the disorder. However, certain diagnostic criteria – chief among which is hyperandrogenism - have been used to delineate the various disease forms into recognizable phenotypes (9,13,63). The Rotterdam classification – reputed to exhibit the most pcos-inclusive diagnostic yardstick due to the exclusion of only the presence of non-hyperandrogenic disorders to make a pcos diagnosis - describes four different variants. Phenotype 1, which corresponds to the classical (full blown) pcos phenotype, should have biochemical and/or clinical hyperandrogenism,

oligo/anovulation, and radiological polycystic ovarian morphology. Phenotype 2 (non-polycystic ovary phenotype) consists of clinical and/or biochemical hyperandrogenism and oligo/anovulation. Phenotype 3 (non-androgenic phenotype) is made of oligo/anovulation and radiological polycystic ovarian morphology; and phenotype 4 (ovulatory phenotype) features comprise biochemical and/or clinical hyperandrogenism and radiological polycystic ovarian morphology (68,69). The National Institute of Health (1990) pcos-diagnostic criteria which require exclusion of hyperprolactinaemia in addition to hyperandrogenism invariably prescribe a pcos variant corresponding to Rotterdam's phenotype 2 for which polycystic ovarian morphology is non-essential. So far, the Androgen Excess Society (2006) diagnostic criteria remain the most stringent requiring the exclusion of concurrent hyperandrogenic disorders e.g., hyperprolactinemia, thyroid hormone imbalance and severe insulin resistance syndromes, and the presence of either clinical hyperandrogenism or hyperandrogenemia plus either oligo/anovulation or ultrasonographic polycystic ovarian morphology (64,66,67).

## CASE REPORT

Our client is a 25-year-old married Nigerian woman who presented herself for a medical consultation September 2023 with a history of irregular and infrequent menstrual cycles which started way back to her menarche at the age of 13 years. Her menstrual cycles which often came once in 3 months and lasted about 5 days then, were with normal flow but associated with severe dysmenorrhea and heart palpitations. At the age of about sixteen years, due to worsening palpitations, she undertook a Cardiological/Gynaecological consultations from which a provisional diagnosis of polycystic ovary syndrome was made based on her personal and gynaecological history; clinical findings of male pattern of chest/abdominal hair distribution; biochemical findings of raised absolute luteinizing hormone (LH) values (about 20 mIU/microL) and increased LH/FSH ratio; and an ultrasonographic finding of polycystic ovaries. Consequently, she was placed on combined oral contraceptive pills (COCs). Following this, her heart palpitations abated and for the next 3 months her menses became more regular but heavy due to COCs-induced synchronised menstrual cycles and withdrawal bleeding, even though her premenstrual and menstrual pains persisted. However, after this period, the heavy menstrual patterns took a twist; this time the COCs no longer achieved the expected withdrawal menstrual bleedings. Desiring to re-establish the lost cyclic withdrawal vaginal bleedings, different COCs were subsequently tried embarked by her doctors for about 3 years with little success. Then this period of nil or scanty menstruation was then followed by an episode of a very heavy thirteen-day-long blood clots-laden menstrual bleeding (menorrhagia). Following this, her subsequent menstrual flows would worsen - experiencing protracted menorrhagia and menstrual bleeding lasting periods longer than a month in several of her menstrual cycles. This phase of her pcos clinical course lasted for another 2-3 years till early 2020. Again, to curtail the protracted heavy menstrual bleeding, she was switched from the combined oral contraceptive pills she was all along placed on to a treatment regime alternating between oral and injectable progesterone-only steroid treatments. Again, there seemed to be a little initial clinical improvement in the heaviness and duration of her menstrual bleeding for the first three cycles of the new progestin-only hormonal therapy. However, after this initial improvement, her menstrual

bleeding got heavier and longer and would continue even while on steroid drugs. The bleedings were often associated with severe dysmenorrhea. The client also volunteered that due to the protracted nature of her menstrual symptoms she started having subjective mixed feelings of internal heat, tension, apprehension, unhappiness, and low moods over the past 2-3 years before her presentation to us. Other aspects of her social history reveal her meals during her teenage years consisted of the typical African diets with occasional indulgence in junk foods. Significantly, her mother does not but a maternal aunt of hers does have a history and features suggestive of pcos and/or infertility. This was her clinical state until she sought our medical consultation in August 2023. At presentation, she was an anxious-looking, slim, averagely tall young woman with generally normal vital signs except for a moderate pallor and a rapid full radial pulse (a finding likely to have resulted from her protracted bleeding and the attendant anaemia). Significantly, there was a finding of a male pattern hair distribution on her chest and abdomen but there was no acne. There was also neither eye proptosis/exophthalmos or fidgetiness (to suggest hyperthyroidism) nor was there lactation from self-pressed breast nipples (to suggest hyperprolactinaemia).

Serum luteinizing hormone (LH) level was 29.75 mIU/mL (Normal range: 2.12-10.89 mIU/mL) and Follicle stimulating hormone (FSH) level was: 3.4 mIU/mL (Normal range: 1.4-9.4 mIU/mL); LH/FSH ratio, 8.75 (normal ratio range 1.00-2.00 ratio). Progesterone level, 0.39 ng/ml (at the lowest end of the normal range: 0.0 - 12.8 ng/ml); serum insulin of 140.00 picomol/L (range: 12-150 ng/ml). Serum cortisol was 29.51 pmol/L (Normal range: 50-230). Serum testosterone level was slightly elevated to 0.93 ng/ml (Normal value <0.80 ng/ml). serum lipid and sugar levels were all within normal values. Serum triiodothyronine of 0.82 ng/ml (Normal range: 0.69-2.15 ng/ml). An ultrasonographic scan revealed multiple well-defined peripheral intraovarian cysts bilaterally. Her psychological score on Beck Depression Inventory (BDI) was 22 (68).

Based on the history, BDI score, clinical, biochemical, and radiological findings, a diagnosis of contraceptive hormones resistant-pcos (Rotterdam phenotype 1) with mild/moderate anaemia, clinical depression, and borderline hyperinsulinaemia was made. She was then placed on haematinics to correct the menorrhagia-induced anaemia. In addition to this, she was tailed off all steroid hormonal medications while she was advised to also continue feeding on locally sourced low table sugar/saturated fat, high fibre, protein, vegetable, and fruit diets she had been placed on from her previous medical consultations. This management was maintained for the following 6-8 weeks, with the haematological and nutritional status improving impressively, but the main pcos symptom, irregular and protracted menstrual bleeding not showing any alleviation. Owing to the severity of her symptoms and their seeming resistance to steroid contraceptive treatments coupled with the reported relative safety, nutritional uses, and anti-pcos potential of MS and TC extracts (69-73) she was then started by our team on a twice daily hot water decoctions of TC brown (fallen) leaves obtained from neighbourhood TC trees (dried and pulverized into fine powder) and MS (Spear mint) leaf tea obtained from nearby commercial stores. Specifically, the regime comprises 2.5 g of MS and one (1) teaspoonful of TC dry leaf powder separately decocted in about 250 ml of hot water and allowed to cool off before being orally taken in the

mornings and evenings for several weeks/months - so long as the symptoms remained. However, within a few days of initiating this herbal combination, the client notified the team she perceived a reduction in the volume (heaviness) of her vaginal bleeding. And about two weeks after the start of the herbal therapy she submitted the bleeding had eventually ceased. The observed clinical progress in her menstrual symptoms was sustained for the following several months - with the cycles becoming progressively more regular and the menstrual flow getting lighter and shorter with each successive menstrual cycle until her cycle lengths stabilized at 26 and 33 days and her monthly menstrual flows at 5-6 days. Over the same period, there was both subjective (self-reporting) and objective (observable) improvement in her moods - scoring less than 10 on a repeat BDI scoring within 3 months of being placed on the MS+TC leaf therapy. The client now felt happier, less tense, less worried, with little or no internal heat. This clinical progress was also mirrored by improvement in most of her biochemical indices. However, there was no commensurate improvement in her hirsutism and dysmenorrhea. This was her clinical picture until around mid-2024 when she informed us (her doctors) that her next menstrual period being expected around May ending 2024 was unusually delayed - even if the little residual fluctuations in her now largely normalised menstrual cycles/flows were to be considered. This delay was also associated with bloating, fatigue, nausea and other symptoms suggestive of clinical malaria - for which she was treated. However, there was no improvement in this latest set of complaints despite adequate antimalaria treatment. This realization, coupled with the unexpected delay of her menses in view of the overall clinical improvement recorded in her reproductive symptoms, prompted a urine pregnancy test, which turned out positive, and a subsequent uterine ultrasonographic scan, which confirmed a six-week old intrauterine viable gestation. However, despite her impressive overall clinical progress, our client's hirsutism remained largely unresolved. To address this, a fresh study by our team to assess the specific anti-hirsutism efficacy of different oil extracts of MS and TC is already ongoing.

## DISCUSSION

Both the case report and the reviewed literature in our study highlight the following facts. Firstly, pcos prevalence is high and under-reported. The high prevalence may be related to the increasing adoption of western-style diets (8-10). Secondly, despite the high prevalence and probability of it increasing further, there is still no streamlined drug treatment for pcos, making treatment approaches largely symptomatic (11,12). This therapeutic conundrum may be related to the fact that the disease aetiopathogenesis is still not completely deciphered (6-8). Thirdly, oral steroids and other synthetic drugs currently used as first/default treatment options have significant efficacy questions and intolerable adverse drug reactions - some of which are being reported in this study (74-77). Lastly, despite widespread anecdotal and scientific reports of the potential efficacy of phytotherapy in pcos management, medicinal plant extracts and natural products are still being both under-investigated and under-utilised for this syndrome (78-81). The foregoing is well illustrated by the impressive anti-pcos and even positive mood-modulatory effects MS and TC leaf extract decoctions exhibited in this report. The common pcos clinical indications for which combined oral contraceptives (COCs) are still considered first therapeutic option include menstrual

irregularity, acne, and hirsutism. COCs are believed to be efficacious for these hyperandrogenaemia-driven indications by a dual mechanism of coupling decreasing ovarian androgen production and plasma free androgen with increasing hepatic sex-hormone binding globulins (SHBGs) (82). Two other symptoms present in our case report for which COCs have been scheduled to be effective are menorrhagia and dysmenorrhea (82,83). However, this client claimed none of these symptoms recorded any clinical improvement in the 2- 4 years she was on oral contraceptives. Reports of complications arising from the use of COCs that are not different from the symptoms they are meant to cure are rampant in clinical practice. Apart from those stated in this presentation, there is a plethora of other potential long-term risks to which this and other pcos cases placed on COCs are exposed. These include obstetric (abortions, intrauterine fetal deaths, preeclampsia, macrosomia, increased fetal congenital anomalies, etc.), renal (failures, hypertension), cancers, metabolic (type 2 diabetes mellitus, obesity, hyperlipidemia, gestational diabetes mellitus), neuropsychiatric (anxiety, depression), and cardiovascular (hypertension, cardiomyopathies, coronary heart disease) complications (84–87). Other steroidal and non-steroidal anti-androgen drugs used in managing features of hyperandrogenism like hirsutism, acne, oligo/anovulation etc. such as metformin, spironolactone, cyproterone acetate, bicalutamide, and flutamide, apart from poor efficacy, are also saddled with intolerable adverse effects like hepatotoxicity, ketoacidosis, nephrotoxicity, hyperkalaemia, cardiotoxicity, etc. (88–92).

Despite the above-stated attendant risks associated with their use, the fact that steroid hormones are still regarded as default prescriptions for hyperandrogenaemia symptoms is a clear indication of the unmet therapeutic gap existing in the management of pcos cases and a pointer to the urgent need to find alternative/complimentary therapeutic approaches that are natural, safer, and acceptable for both effective pcos case treatment and prevention of the highlighted potential long-term risks. Phytotherapy has always provided a promising alternative to synthetic drugs in the treatment of several disease classes: the ongoing achievement of progressive symptom relief by MS and TC leaf extracts over a short period compared to that of COCs in this presentation is an indication that pcos is not an exemption to the broad efficacy spectrum of phytotherapeutic agents(93,94). The wide therapeutic latitude of botanicals is viewed to derive from their inherent possession of diverse potent disease-fighting macromolecules, including particularly, antioxidant compounds(95). In respect to pcos, anecdotal, preclinical and clinical reports indicate several medicinal plant extracts e.g., *Cnidium officinale*(96,97), *Leonurus japonicus*(98,99), *Berberis vulgaris* L.(100) , *Angelica sinensis*(101), *Angelica gigantis*(102) and natural products e.g., berberine (103,104), folate (105), inositol (106–108), Luteolin and luteolin-7-methyl ether (109), paeoniflorin (110), apigenin (111,112), and catechins (113) have been effectively used to alleviate pcos symptoms -especially those related to cellular oxidative stress. As earlier highlighted, chronic systemic low-grade cellular oxidative stress is a hallmark of chronic inflammatory diseases including pcos (27,29). Relatedly, it is often viewed that their intrinsic antioxidant property to reverse/slow down cellular oxidative processes may explain why strategies involving the use of medicinal plant extracts and diets - which are inherently antioxidant, are being increasingly recognized as potential alternative treatment modalities for pcos (31,32). What more,

antioxidant activity (34,35) is one of the best known biological properties of the two medicinal plant extracts being reported for their effective control of pcos menstrual symptoms in this study.

The first of the two medicinal plants in this report, *Mentha spicata* L. (MS) (Common name: spearmint), which has been investigated and reported for its ethnomedicinal and ethnopharmacological anti-pcos activities (70,114–116), is a creeping rhizomatous, strongly aromatic perennial hybrid herb native to Europe, the Americas, and Asia but now widely cultivated in virtually all cultures, often as a back garden flowery plant. This Lamiaceae family medicinal plant that can grow up to 100 cm high has over a hundred synonyms including *Mentha condensis* L., *Mentha viridis* (L.), *Mentha cordifolia*, *Mentha balsamea* Rchb., *Mentha aquatica* L., *Mentha laevigata* Willd., *Mentha spicata* L., *Mentha brevispicata* Lehm., and *Mentha piperita*, *Mentha crispa* var. *crispata*, *reticulata*, and *Mentha glabra* Mill. (117,118). Essential oils, decoctions, infusions, and powders from different parts of spearmint have been traditionally administered to alleviate peptic ulcer disease, low libido, headache, fatigue, fevers, sore throat, diabetes mellitus, chest pain and infections, kidney ailments, asthma, colds, and flatulence (119,120). Oils and extracts from leaf, stem, flowers, and roots have been shown to be antifungal, insecticidal, antibacterial, antiparasitic, analgesic and antipyretic, anti-inflammatory, hypoglycemic, antioxidant, antiproliferative, and antidiuretic (121–125). As pointed out above, scientific reports on MS extracts indicate they are effective in managing female reproductive health diagnoses such as infertility, hyperandrogenaemia, menorrhagia, and dysmenorrhea in experimental animal studies and human clinical trials (78,79). Studies have further shown extracts of MS alone, or in combination, ameliorated ovarian follicular dysgenesis and hyperandrogenesis in rats (78,116), decreased significantly ovarian cystic folliculogenesis, adrogenaemia and oestrogenaemia, and increased the number and quality of ovarian follicles across all stages of development in rats (116,126).

Human studies have also reported MS extracts orally or topically administered for various periods in pcos women caused significant free and total testosterone reduction, hirsutism severity reduction, and improved ovarian function and morphology (79,127). It is worth to note that in a study by Bahl, A. S. (2020)(128), topically administered MS extract was so effective in the MS-treatment group that over 36% of enlisted pcos women exposed to MS oil, compared to zero percent in the placebo-exposed group, overcame their existing infertility to achieve conception within six-month of post-trial follow up (127). It is our opinion that the impressive pro-conception report of MS oil application in pcos women appears to be surpassed by our finding whereby our client did achieve conception right while still on MS and TC therapy. The pro-conception inclination of MS oil, and MS+TC hot water decocts being highlighted in these studies may also be related to their potent antioxidant activity. In fact, the overall anti-pcos effect of MS extracts on the back of their antioxidant and anti-androgenic activity is viewed to be due to their rich possession of (116,129) of bioactive essential oils and terpenes made up of diverse phytochemicals including carvone, dihydrocarvone, trans-carveol, menthone,  $\beta$ -caryophyllene, carvacrol, germacrene D, limonene, menthol, etc.(130,131).

*Terminalia catappa* (TC), the second of the two medicinal plants combined in the herbal treatment in this report, is a tropical monoecious flowering plant of the Combretaceae family, initially native to Africa and Southeast Asia but now cultivated in most parts of the world (132,133). This medicinal and nutritional plant has several synonyms such as *Terminalia catappa* var. *chlorocarpa* Hassk., *Terminalia catappa* var. *macrocarpa* Hassk., *Terminalia chebula*, *Terminalia catappa* var. *subcordata* (Humb. & Bonpl. ex Willd.) DC., *Terminalia bellirica*, *Terminalia burmanica* King ex Prain., *Terminalia badamia* sensu Tul., *Terminalia catappa* var. *pubescens* Kurz, *Terminalia catappa* var. *rhodocarpa* Hassk., etc., and common names such as 'fruit', 'tropical almond', 'autaraa', 'kamani haole', 'kaukauriki', 'Indian almond', 'talie', 'talise', 'tavola', 'tivi', in different cultures. This plant which can grow up to 35 metre tall has large 15-25 cm x 10-14 cm ovoid, dark green, and glossy/leathery leaves and 5-7 cm long x 3-5.5 cm green, yellow, or red (depending on degree of ripeness) succulent fruits – each containing a single shelled edible seed (134–137). Although TC extracts have been reported effective for diverse ethnobotanical and ethno pharmacological indications, our literature search revealed little or no scientific studies on the anti-pcos efficacy of extracts obtained from TC unlike those from MS, except for two of its synonyms – *Terminalia chebula* and *Terminalia bellirica* (138–140). The several therapeutic indications for which extracts of the kernel, leaves, stem bark, seed, fruits, roots, and flowers of TC have been ethnomedicinally used include male sexual disturbances (141), rheumatic arthritis, skin rashes/dermatitis, fevers (81,142,143), depression (80), hypertension (144), inflammatory and oxidative stress-related immune dysfunction (145), inflammatory and cancerous liver diseases (146), and to enhance longevity (147).

Similarly, the diverse pharmacological activities already reported for root, stem bark, fruit, seed, and leaf extract of TC include antidepressant (72,80,148), anxiolytic (71,72,149), anti-hypertensive (150–153), aphrodisiac (154,155), antilipidaemic (156,157), and analgesic, antipyretic, and inflammatory (158–160). Other reported activities of TC extracts include hepatoprotective (161,162), interleukin-6 (IL-6) gene expression-suppressant (160), antihyperglycaemic (163,164), anti-parasitic and antimicrobial (165–169), immunomodulatory (170,171), wound-healing and anti-ageing (172,173), anticarcinogenic/antimutagenic (174–176), and neuroprotective (177). These biological activities of TC are believed to derive from the widely reported potent antioxidant property (137,178,179) of the large arrays of bioactive phytochemicals, such as flavonoids, beta-carotene, palmitoleic-acid, ursolic acid, castalagin, leucocyanidin, Linoleic-acid, eugenigrandin A,  $\beta$ -sitosterol, punicalagin, betulinic-acid, tannin, alkaloids, gallic-acid, corilagin, daucosterol, ellagic-acid, niacin, saponins, brevifolin-carboxylic-acid, steroids, oleic-acid, ellagic-acid, tercatanin, oleanolic-acid, oxalic-acid, cardiac glycosides, grandinin, 2, 3, 23-trihydroxyurs-12-en-28-oic acid, myristic-acid, palmitic-acid, Corilagin, cyanidin-3-glucoside, coumarins, riboflavin, pentosans, tannin. Magnesium, potassium, vitamin A, phosphorus, and many more, present in various parts of this medicinal plant (180–186). Taken together, our literature search implies antioxidant activity appears to be a key mechanism by which MS, TC, other medicinal plant extracts, and even some non-herbal treatments effective in pcos management exert their anti-pcos potency. This view is premised on the understanding of the central role systemic cellular inflammation and oxidative stress

from diverse epigenetic, genetic, and environmental stimuli play in triggering systems-wide tissue steroidogenic activity of enzymes (dehydrogenases (3 $\beta$ -hydroxysteroid dehydrogenase type-1), reductases, hydroxylases) and other pro-androgenic proteins (steroidogenic acute regulatory proteins (StAR) and Steroidogenic Factor-1(SF-1)) towards hyperandrogenaemia and hyperandrogenism – the key biochemical features underpinning pcos (18–26,28,187–192). Indeed, antioxidant effect is likely the main factor behind the reported potential therapeutic benefits of vegetarian, organic or ketogenic dieting on one hand (193,194), and on the other, lack of antioxidant activity of high carbohydrate and low fibre dieting is behind the preponderant association of Advanced Glycation End-products (AGEs)-dominated dieting with pcos pathogenesis (195,196). Similar rationale lies behind the advisory to pcos victims and predisposed persons to scale down on AGEs-related foods and cultivate more pro-organic diets comprising fruits, vegetables, whole grains, and healthy plant and animal proteins (197–200). This is necessary because overweight resulting from habitual indulgence in AGEs, overfried/processed foods, cigarette smoking, refined sugar, and sedentary routines promotes cellular oxidative stress in the long term (195–200). Up taking antioxidant effect from herbal, dietary and/or kinetic interventions to mitigate or reverse oxidative stress and potentially, the pcos process and the associated insulin resistance is therefore recommended (201–203).

Preclinical studies suggest high-dose folate supplementation is a potential anti-pcos therapeutic intervention in fetuses of pcos mothers (204). It is of interest to note that the main mechanistic thrust of this epigenetic treatment is the electron (methyl group) donation-fuelled antioxidant potency of the natural folates, their physiological form, folic acid, and their basic antioxidant pharmacophore, 4-hydroxy-2,5,6-triaminopyrimidine (205–208). These therapeutic agents have been shown to possess potent free radical scavenging and ferric iron reducing activity (205,208). Research indicates the antioxidant activity of reduced folates and folic acid most likely accounts for their therapeutic efficacy for diverse therapeutic indications such as metabolic, neoplastic, neurodegenerative, cardiovascular, haematological, reproductive, etc. (49,209–211). Animal studies have shown the antioxidant activity of high-dose folate/folic acid supplementation ameliorated circulatory overload-induced left cardiac ventriculopathy and collagen loss in rat, mitigated negative foetal epigenetic impact, and reduced radiation-induced functional and morphological ovarian damage in mice (205, 207,212).

Similarly clinical studies indicate the antioxidant activity of high-dose folate/folic acid supplementation improved patients' clinical indices across a broad disease spectrum. For instance, chronic 10-mg daily folic acid supplementation ameliorated both carotid arteriosclerotic endothelial dysfunction and intima-media mitogenesis (213,214). Furthermore, randomised, double-blind, placebo-controlled studies in overweight and obese pcos women show a 5mg daily folate supplementation for various periods had positive impact on their inflammatory factors and oxidative stress biomarkers, improved glucose/lipid profiles/tissue insulin sensitivity, and ameliorated hyper homocysteinaemia (210–212,215,216). Evidence indicates the antioxidant activity of folates/folic acid is shared by most antioxidant compounds - herbal and natural products such as myo-inositol (217,218), alpha( $\alpha$ )-lipoic acid

(ALA,  $\alpha$ LA) (219), and the sirtuins – a group of endogenous regulatory proteins that are pro-life nicotinamide adenine dinucleotides (NAD<sup>+</sup>)-dependent class III histone deacetylases (NHDACs) (220). Both myo-inositol/inositol derivatives (221–226) and  $\alpha$ -lipoic acid/lipoate derivatives (219,227–229) have been implicated in the modulation of signal transduction, neuroinflammation, neuroendocrine signalling – including insulin sensitivity and lipogenesis, cellular reactive oxygen species generation, and regulation of ovarian cycle, folliculogenesis, and steroidogenesis. Thus, their individual and shared anti-inflammatory, antioxidant, insulin-mimicking, and ovarian function regulatory profiles – and knowing the important roles chronic inflammation, tissue insulin insensitivity, cellular oxidative, ovary dysfunction and dyssteroidogenesis play in pcos disease process make myo-inositol (217,222,230,231) and ALA (227–229,232–236) individually and synergistically (237–240) potentially effective in its management.

The sirtuins, of which the human homologous form is sirtuin1, SIRT1, are a group of stress- and calorie-sensitive proteins (genes) whose catalytic regulatory influence on cell metabolism is premised on acetylation-based posttranslational modification of other metabolic proteins (241–246). Acting as cellular calorie/energy sensors, these endogenous antioxidant proteins (247–249) are known to play key roles in diverse biological processes like calorie restriction response, cell cycle progression, transcription, chromatin remodelling, cell proliferation, ovarian granulosa cell and follicle growth, development and steroidogenesis, tumorigenesis, apoptosis, insulin signalling, DNA repair, senescence and ageing, cellular and mitochondrial inflammation, oxidative stress and reactive oxygen species scavenging, circadian rhythm, cell replication and cycle (242,250–257). Like myo-inositol and ALA, these highlighted activities appear to confer significant anti-pcos efficacy on sirtuins, as pcos amelioration in both animal and human studies has been reported for them (258–262). It is apt to mention that the fact that inositol and ALA and their derivatives are products of natural/botanical sources (263,264) like MS and TC on one hand, and the fact that phenolic phytochemicals (e.g., resveratrol, naringenin, etc.) have been shown to activate the sirtuin (SIRT1)-based anti-pcos activity (259,260,262,265,266) in previous studies on the other, may imply the observed anti-pcos effects of MS and TC hot water decoctions being reported in this study may be partly due to their possession of yet to be identified antioxidant inositol and ALA derivatives as well as modulation of the free radicals scavenging SIRT1-linked anti-pcos activity. Perhaps the lack of high antioxidant activity, unlike MS, TC, the folates, ALA, myo-inositol, and the sirtuin proteins, by the existing synthetic anti-pcos drugs like contraceptive hormones, metformin, spironolactone, cyproterone acetate, bicalutamide, and flutamide – may partly account for their perceived low anti-pcos efficacy and high toxicity (83,86–92). Additional advantage the use of phytotherapy may have over synthetic pharmacotherapy in the treatment of pcos is buttressed in our study by the subjective and objective alleviation of our client's neuro-psychological symptoms within weeks following twice daily oral administration of MS and TC hot water decoctions. The mood lifting activity of these extracts, which may be closely related to their possession of potent antioxidant macromolecules such as ursolic acid, gallic acid, carvone, menthone, etc. (118,130,135,137,143,267), is a finding that should not be overlooked but further probed, developed and appropriated.

Anxiety and depression are common features of pcos and related reproductive disorders. Already, findings from previous studies support the mood-alleviating activity of extracts from both MS and TC. This finding is supported by previous studies reporting significant antioxidant and antidepressant effects from MS essential oil, MS and TC phenolic compounds, and TC hydrolysable tannins (80,129,148,267–269). The inherent capacity of phytotherapeutic agents, unlike the synthetic drugs, to be effective across different body systems and pathophysiological pathways is well show-cased in the current report by the herbal extracts concurrently mitigating our client's mood and reproductive symptoms. This fact is yet another comparative strength of phytotherapy over synthetic pharmacotherapy. Does psychotherapy have a role in pcos treatment? Certainly yes, psychological counselling or behavioural therapy will be helpful in the clinical progress of these cases, especially when combined with the standard pharmacological treatment modalities. Many pcos women often are overweight or obese, prone to spells of low esteem, anxiety, depression, and sometimes, rejection – most of which psychotherapy will not only help to alleviate but also engender compliance to anti-pcos medications (270–272). Psychotherapy is also useful in the initiation and maintenance of positive lifestyle changes -including right kinetic, dietary, and psychosocial choices that will ultimately achieve and sustain the overall therapeutic goal (273–275).

## CONCLUSION

This report highlights pcos high prevalence, existing anti-pcos therapeutic gap due to non-existent standard pcos treatments and the efficacy/toxicity challenges with the existing synthetic anti-pcos agents. The study also reports the effectiveness of a polyherbal treatment in alleviating pcos and associated mood symptoms. This outcome points to unexploited potential in phytotherapy and justifies the traditional uses of these medicinal plant extracts to remediate reproductive and neuropsychiatric symptoms. It is necessary to further develop phytomedicine to meet modern-day health needs.

## REFERENCES

1. Azziz R, Adashi EY. Stein and Leventhal: 80 years on. *Am J Obstet Gynecol.* 2016 Feb 1;214(2):247.e1-247.e11.
2. Leventhal ML. The Stein-Leventhal syndrome. *Am J Obstet Gynecol.* 1958 Oct 1;76(4):825–38.
3. Pesonen S, Timonen S, Mikkonen R. Symptoms and etiology of the stein-leventhal syndrome. *Acta Endocrinol (Copenh).* 1959 Mar;30(3):405–23.
4. David A. Ehrman RBBRLR. Polycystic ovary syndrome as a form of functional ovarian hyperandrogenism due to dysregulation of androgen secretion. *Endocr Rev.* 1995 Jun 1;16(3):322–53.
5. Wolf WM, Wattick RA, Kinkade ON, Olfert MD. Geographical prevalence of polycystic ovary syndrome as determined by region and race/ethnicity. Vol. 15, *International Journal of Environmental Research and Public Health.* MDPI; 2018.
6. Pathak G, Nichter M. Polycystic ovary syndrome in globalizing India: An ecosocial perspective on an emerging lifestyle disease. *Soc Sci Med.* 2015;146.
7. Goodarzi MO, Azziz R. Diagnosis, epidemiology, and genetics of the polycystic ovary syndrome. Vol. 20, *Best*

- Practice and Research: Clinical Endocrinology and Metabolism. 2006.
8. Bednarska S, Siejka A. The pathogenesis and treatment of polycystic ovary syndrome: What's new? Vol. 26, *Advances in Clinical and Experimental Medicine*. 2017.
  9. Parker J, O'Brien C, Hawrelak J, Gersh FL. Polycystic Ovary Syndrome: An Evolutionary Adaptation to Lifestyle and the Environment. Vol. 19, *International Journal of Environmental Research and Public Health*. MDPI; 2022.
  10. Merkin SS, Phy JL, Sites CK, Yang D. Environmental determinants of polycystic ovary syndrome. Vol. 106, *Fertility and Sterility*. 2016.
  11. Dumesic DA, Oberfield SE, Stener-Victorin E, Marshall JC, Laven JS, Legro RS. Scientific statement on the diagnostic criteria, epidemiology, pathophysiology, and molecular genetics of polycystic ovary syndrome. Vol. 36, *Endocrine Reviews*. 2015.
  12. Barrett ES, Sobolewski M. Polycystic ovary syndrome: Do endocrine-disrupting chemicals play a role? *Semin Reprod Med*. 2014;32(3).
  13. Parker J. Pathophysiological Effects of Contemporary Lifestyle on Evolutionary-Conserved Survival Mechanisms in Polycystic Ovary Syndrome. Vol. 13, *Life*. 2023.
  14. Jiang L Le, Xie JK, Cui JQ, Wei D, Yin BL, Zhang YN, et al. Promoter methylation of yes-associated protein (YAP1) gene in polycystic ovary syndrome. *Medicine (United States)*. 2017;96(2).
  15. Shukla P, Mukherjee S. Mitochondrial dysfunction: An emerging link in the pathophysiology of polycystic ovary syndrome. Vol. 52, *Mitochondrion*. 2020.
  16. Zhao S, Tian Y, Gao X, Zhang X, Liu H, You L, et al. Family-based analysis of eight susceptibility loci in polycystic ovary syndrome. *Sci Rep*. 2015;5.
  17. Piltonen TT, Giacobini P, Edvinsson Å, Hustad S, Lager S, Morin-Papunen L, et al. Circulating antimüllerian hormone and steroid hormone levels remain high in pregnant women with polycystic ovary syndrome at term. *Fertil Steril*. 2019;111(3).
  18. Kim JJ, Choi YM, Hong MA, Hwang SS, Yoon SH, Chae SJ, et al. Phosphatidylinositol 3-kinase p85 $\alpha$  regulatory subunit gene Met326Ile polymorphism in women with polycystic ovary syndrome. *Human Reproduction*. 2009;24(5).
  19. Maliqueo M, Sundström Poromaa I, Vanky E, Fornes R, Benrick A, Åkerud H, et al. Placental STAT3 signaling is activated in women with polycystic ovary syndrome. *Human Reproduction*. 2015;30(3).
  20. Zhou S, Tang X, Chen HZ. Sirtuins and Insulin Resistance. *Front Endocrinol (Lausanne)*. 2018 Dec 6;9.
  21. Zilocchi M, Colugnat I, Lualdi M, Meduri M, Marini F, Corasolla Carregari V, et al. Exploring the Impact of PARK2 Mutations on the Total and Mitochondrial Proteome of Human Skin Fibroblasts. *Front Cell Dev Biol*. 2020;8.
  22. Huo Y, Ji S, Yang H, Wu W, Yu L, Ren Y, et al. Differential expression of microRNA in the serum of patients with polycystic ovary syndrome with insulin resistance. *Ann Transl Med*. 2022;10(14).
  23. Kyeong Song D, Sung YA, Lee H. The role of serum microRNA-6767-5p as a biomarker for the diagnosis of polycystic ovary syndrome. *PLoS One*. 2016;11(9).
  24. MA SG, M TS. Metabolic dysfunction in polycystic ovary syndrome: Pathogenic role of androgen excess and potential therapeutic strategies. *Mol Metab*. 2020;35.
  25. Mohammadi M. Oxidative stress and polycystic ovary syndrome: A brief review. *Int J Prev Med*. 2019;10(1).
  26. Siddiqui S, Mateen S, Ahmad R, Moin S. A brief insight into the etiology, genetics, and immunology of polycystic ovarian syndrome (PCOS). Vol. 39, *Journal of Assisted Reproduction and Genetics*. 2022.
  27. Zuo T, Zhu M, Xu W. Roles of oxidative stress in polycystic ovary syndrome and cancers. Vol. 2016, *Oxidative Medicine and Cellular Longevity*. 2016.
  28. Di Berardino C, Peserico A, Capacchietti G, Zappacosta A, Bernabò N, Russo V, et al. High-Fat Diet and Female Fertility across Lifespan: A Comparative Lesson from Mammal Models. Vol. 14, *Nutrients*. 2022.
  29. Liu Y, Liu H, Li Z, Fan H, Yan X, Liu X, et al. The Release of Peripheral Immune Inflammatory Cytokines Promote an Inflammatory Cascade in PCOS Patients via Altering the Follicular Microenvironment. *Front Immunol*. 2021;12.
  30. Bahia L, Aguiar LG, Villela N, Bottino D, Godoy-Matos AF, Geloneze B, et al. Relationship between adipokines, inflammation, and vascular reactivity in lean controls and obese subjects with metabolic syndrome. *Clinics*. 2006;61(5).
  31. Yang W, Bai X, Luan X, Min J, Tian X, Li H, et al. Delicate regulation of IL-1 $\beta$ -mediated inflammation by cyclophilin A. *Cell Rep*. 2022;38(11).
  32. Tornatore L, Thotakura AK, Bennett J, Moretti M, Franzoso G. The nuclear factor kappa B signaling pathway: Integrating metabolism with inflammation. Vol. 22, *Trends in Cell Biology*. 2012.
  33. Bremer AA, Miller WL. The serine phosphorylation hypothesis of polycystic ovary syndrome: a unifying mechanism for hyperandrogenemia and insulin resistance. Vol. 89, *Fertility and Sterility*. 2008.
  34. Unluhizarci K, Karaca Z, Kelestimur F. Role of insulin and insulin resistance in androgen excess disorders. *World J Diabetes*. 2021;12(5).
  35. Maiorino MI, Bellastella G, Giugliano D, Esposito K. From inflammation to sexual dysfunctions: a journey through diabetes, obesity, and metabolic syndrome. Vol. 41, *Journal of Endocrinological Investigation*. 2018.
  36. Dube S, Errazuriz I, Cobelli C, Basu R, Basu A. Assessment of insulin action on carbohydrate metabolism: Physiological and non-physiological methods. Vol. 30, *Diabetic Medicine*. 2013.
  37. Soldani R, Cagnacci A, Yen SSC. Insulin insulin-like growth factor I (IGF-I) and IGF-II enhance basal and gonadotrophin-releasing hormone-stimulated luteinizing hormone release from rat anterior pituitary cells in vitro. *Eur J Endocrinol*. 1994;131(6).
  38. Krüger M, Kratchmarova I, Blagoev B, Tseng YH, Kahn CR, Mann M. Dissection of the insulin signaling pathway via quantitative phosphoproteomics. *Proc Natl Acad Sci U S A*. 2008;105(7).
  39. Chan O, Inouye K, Akirav E, Park E, Riddell MC, Vranic M, et al. Insulin alone increases hypothalamo-pituitary-adrenal activity, and diabetes lowers peak stress responses. *Endocrinology*. 2005;146(3).
  40. Messinis IE, Messini CI, Dafopoulos K. Novel aspects of the endocrinology of the menstrual cycle. Vol. 28, *Reproductive BioMedicine Online*. 2014.

41. Nestler JE, Powers LP, Matt DW, Steingold KA, Plymate SR, Rittmaster RS, et al. A direct effect of hyperinsulinemia on serum sex hormone-binding globulin levels in obese women with the polycystic ovary syndrome. *Journal of Clinical Endocrinology and Metabolism*. 1991;72(1).
42. Herman R, Sikonja J, Jensterle M, Janez A, Dolzan V. Insulin Metabolism in Polycystic Ovary Syndrome: Secretion, Signaling, and Clearance. Vol. 24, *International Journal of Molecular Sciences*. 2023.
43. Magoffin DA. Evidence that luteinizing hormone-stimulated differentiation of purified ovarian thecal-interstitial cells is mediated by both type I and type II adenosine 3',5'-monophosphate-dependent protein kinases. *Endocrinology*. 1989;125(3).
44. Miller WL, Tee MK. The post-translational regulation of 17,20 lyase activity. *Mol Cell Endocrinol*. 2015 Jun 15;408:99–106.
45. Taylor AE, Mccourt B, Martin KA, Anderson EJ, Adams JM, Schoenfeld D, et al. Determinants of abnormal gonadotropin secretion in clinically defined women with polycystic ovary syndrome. *Journal of Clinical Endocrinology and Metabolism*. 1997;82(7).
46. Cara JF, Rosenfield RL. Insulin-like growth factor i and insulin potentiate luteinizing hormone-induced androgen synthesis by rat ovarian thecal-interstitial cells. *Endocrinology*. 1988;123(2).
47. Mortensen M, Ehrmann DA, Littlejohn E, Rosenfield RL. Asymptomatic volunteers with a polycystic ovary are a functionally distinct but heterogeneous population. *Journal of Clinical Endocrinology and Metabolism*. 2009;94(5).
48. Nie C, Li Y, Qian H, Ying H, Wang L. Advanced glycation end products in food and their effects on intestinal tract. Vol. 62, *Critical Reviews in Food Science and Nutrition*. 2022.
49. De Vos WM, Tilg H, Van Hul M, Cani PD. Gut microbiome and health: mechanistic insights. *Gut*. 2022;71(5).
50. Weidemann A. *Comprehensive Guide to Nutrition in Polycystic Ovarian Syndrome (PCOS)*. Comprehensive Guide to Nutrition in Polycystic Ovarian Syndrome (PCOS). 2019.
51. Padmanabhan V, Song W, Puttabyatappa M. Praegnatio perturbatio—impact of endocrine-disrupting chemicals. Vol. 42, *Endocrine Reviews*. 2021.
52. Ahn C, Jeung EB. Endocrine-Disrupting Chemicals and Disease Endpoints. Vol. 24, *International Journal of Molecular Sciences*. 2023.
53. Ma M, Wang M, Xu F, Hao S. The Imbalance in Th17 and Treg Cells in Polycystic Ovarian Syndrome Patients with Autoimmune Thyroiditis. *Immunol Invest*. 2022;51(5).
54. Luan Y yi, Zhang L, Peng Y qiu, Li Y ying, Liu R xia, Yin C hong. Immune regulation in polycystic ovary syndrome. Vol. 531, *Clinica Chimica Acta*. 2022.
55. Nelson VL, Legro RS, Strauss JF, McAllister JM. Augmented androgen production is a stable steroidogenic phenotype of propagated theca cells from polycystic ovaries. *Molecular Endocrinology*. 1999;13(6).
56. Bhatt MR, Khatri Y, Rodgers RJ, Martin LL. Role of cytochrome b5 in the modulation of the enzymatic activities of cytochrome P450 17 $\alpha$ -hydroxylase/17,20-lyase (P450 17A1). Vol. 170, *Journal of Steroid Biochemistry and Molecular Biology*. 2017.
57. Auchus RJ, Lee TC, Miller WL. Cytochrome b5 augments the 17,20-lyase activity of human P450c17 without direct electron transfer. *J Biol Chem*. 1998 Feb 6;273(6):3158–65.
58. Tee MK, Miller WL. Phosphorylation of human cytochrome P450c17 by p38 $\alpha$  selectively increases 17,20 lyase activity and androgen biosynthesis. *J Biol Chem*. 2013 Aug 16;288(33):23903–13.
59. Narayana S, Ananad C, Kumari NS, Sonkusere S, Babu SVS. Impact of Aromatase Enzyme and its Altered Regulation on Polycystic Ovary Syndrome (PCOS): A Key Factor in Pathogenesis of PCOS. *Indian Journal of Medical Specialities*. 2023;14(4).
60. Chen J, Shen S, Tan Y, Xia D, Xia Y, Cao Y, et al. The correlation of aromatase activity and obesity in women with or without polycystic ovary syndrome. *J Ovarian Res*. 2015;8(1).
61. Piouka A, Karkanaki A, Katsikis I, Delkos D, Mousatou T, Daskalopoulos G, et al. Association of anti-mullerian hormone and adiponectin in normal weight and overweight plus obese women with polycystic ovary syndrome. *Human Reproduction*. 2011;26(SUPPL. 1).
62. Wang H, Li Q, Wang T, Yang G, Wang Y, Zhang X, et al. A common polymorphism in the human aromatase gene alters the risk for polycystic ovary syndrome and modifies aromatase activity in vitro. *Mol Hum Reprod*. 2011;17(6).
63. Rosenfield RL, Ehrmann DA. The Pathogenesis of Polycystic Ovary Syndrome (PCOS): The hypothesis of PCOS as functional ovarian hyperandrogenism revisited. Vol. 37, *Endocrine Reviews*. 2016.
64. Fauser BCJM. Revised 2003 consensus on diagnostic criteria and long-term health risks related to polycystic ovary syndrome. *Fertil Steril*. 2004;81(1).
65. Yilmaz M, Isaoglu U, Delibas IB, Kadanali S. Anthropometric, clinical and laboratory comparison of four phenotypes of polycystic ovary syndrome based on Rotterdam criteria. *Journal of Obstetrics and Gynaecology Research*. 2011;37(8).
66. Fahs D, Salloum D, Nasrallah M, Ghazeeri G. Polycystic Ovary Syndrome: Pathophysiology and Controversies in Diagnosis. Vol. 13, *Diagnostics*. 2023.
67. International evidence-based guideline for the assessment and management of polycystic ovary syndrome - 2023. *Reproductive Endocrinology*. 2023;(69).
68. Cogan AB, Persons JB, Kring AM. Using the Beck Depression Inventory to Assess Anhedonia: A Scale Validation Study. *Assessment*. 2024;31(2).
69. Teede HJ, Misso ML, Costello MF, Dokras A, Laven J, Moran L, et al. Recommendations from the international evidence-based guideline for the assessment and management of polycystic ovary syndrome. *Human Reproduction*. 2018;33(9).
70. Alaei S, Bagheri MJ, Ataabadi MS, Koohpeyma F. Capacity of Mentha spicata (spearmint) Extract in Alleviating Hormonal and Folliculogenesis Disturbances in Polycystic Ovarian Syndrome Rat Model. *J Worlds Poult Res*. 2020;10(3).
71. Umarudeen AM, Magaji MG, Khan F, Abubakar A. Acute Toxicity and Anxiolytic Activity Screening of Hydroalcoholic Leaf Extracts of Bryophyllum pinnatum, Terminalia catappa and Tapinanthus dodoneifolius Growing on Terminalia catappa Tree in MIce. *Int Neuropsychiatr Dis J*. 2023;19(3).

72. Habibullah B, Panezai MA, Kakar MA, Achakzai JK, Kakar AM, Khan NY, et al. Biological studies on leaves of Tropical Almond (*Terminalia catappa*) (A Review). *Eur Acad Res*. 2023;XI(1).
73. Iheagwam FN, Iheagwam OT, Onuoha MK, Ogunlana OO, Chinedu SN. *Terminalia catappa* aqueous leaf extract reverses insulin resistance, improves glucose transport and activates PI3K/AKT signalling in high fat/streptozotocin-induced diabetic rats. *Sci Rep*. 2022;12(1).
74. Palomba S, Santagni S, Falbo A, La Sala GB. Complications and challenges associated with polycystic ovary syndrome: Current perspectives. Vol. 7, *International Journal of Women's Health*. 2015.
75. Sun Y, Xu T, Zhu S, Xu H. Characteristics of adverse drug reactions induced by flutamide and bicalutamide: a real-world pharmacovigilance study using FAERS. *Expert Opin Drug Saf*. 2024;23(3).
76. R.A. B, M.C. M, A.L. S. Contraception choices in women with underlying medical conditions. *Am Fam Physician*. 2010;82(6).
77. J. D, S. G. The teenager with heavy menses. Vol. 10, *Medicine Today*. 2009.
78. Ataabadi MS, Alae S, Bagheri MJ, Bahmanpoor S. Role of essential oil of *Mentha spicata* (spearmint) in addressing reverse hormonal and folliculogenesis disturbances in a polycystic ovarian syndrome in a rat model. *Adv Pharm Bull*. 2017;7(4).
79. Akdoğan M, Tamer MN, Cüre E, Cüre MC, Köroğlu BK, Delibaş N. Effect of spearmint (*Mentha spicata* Labiatae) teas on androgen levels in women with hirsutism. *Phytotherapy Research*. 2007;21(5).
80. Chandrasekhar Y, Ramya EM, Navya K, Phani Kumar G, Anilakumar KR. Antidepressant like effects of hydrolysable tannins of *Terminalia catappa* leaf extract via modulation of hippocampal plasticity and regulation of monoamine neurotransmitters subjected to chronic mild stress (CMS). *Biomedicine and Pharmacotherapy*. 2017;86.
81. Untwal L, Kondawar M. Use of *Terminalia catappa* fruit extract as an indicator in acid-base titrations. *Indian J Pharm Sci*. 2006;68(3).
82. Dumesic DA, Akopians AL, Madrigal VK, Ramirez E, Margolis DJ, Sarma MK, et al. Hyperandrogenism accompanies increased intra-abdominal fat storage in normal weight polycystic ovary syndrome women. *Journal of Clinical Endocrinology and Metabolism*. 2016;101(11).
83. Vrbíková J, Cibula D. Combined oral contraceptives in the treatment of polycystic ovary syndrome. Vol. 11, *Human Reproduction Update*. 2005.
84. Oguz SH, Yildiz BO. An update on contraception in polycystic ovary syndrome. Vol. 36, *Endocrinology and Metabolism*. 2021.
85. NCT01889199. Androgen Excess as a Cause for Adipogenic Dysfunction in PCOS Women. <https://clinicaltrials.gov/show/NCT01889199>. 2013;
86. Manzoor S, Ganie MA, Amin S, Shah ZA, Bhat IA, Yousuf SD, et al. Oral contraceptive use increases risk of inflammatory and coagulatory disorders in women with Polycystic Ovarian Syndrome: An observational study. *Sci Rep*. 2019;9(1).
87. Dr Sameer Sanghvi. Lloyds Pharmacy Online Doctor . 2021. What are the long-term side effects of birth control pills?
88. van der Spuy ZM, Le Roux PA, Matjila MJ. Cyproterone acetate for hirsutism. Vol. 2010, *Cochrane Database of Systematic Reviews*. 2003.
89. Lumachi F, Rondinone R. Use of cyproterone acetate, finasteride, and spironolactone to treat idiopathic hirsutism. *Fertil Steril*. 2003;79(4).
90. Brahm J, Brahm M, Segovia R, Latorre R, Zapata R, Poniachik J, et al. Acute and fulminant hepatitis induced by flutamide: Case series report and review of the literature. *Ann Hepatol*. 2011;10(1).
91. Wang Y, Lipner SR. Retrospective analysis of adverse events with spironolactone in females reported to the United States Food and Drug Administration. *Int J Womens Dermatol*. 2020;6(4).
92. Adriana Stoica R, Simona Ștefan D, Rizzo M, Iulia Suceveanu A, Paul Suceveanu A, Serafinceanu C, et al. Metformin Indications, Dosage, Adverse Reactions, and Contraindications. In: *Metformin*. 2020.
93. Efferth T, Koch E. Complex Interactions between Phytochemicals. The Multi-Target Therapeutic Concept of Phytotherapy. *Curr Drug Targets*. 2010;12(1).
94. Srivastava A, Srivastava P, Pandey A, Khanna VK, Pant AB. Phytomedicine: A Potential Alternative Medicine in Controlling Neurological Disorders. In: *New Look to Phytomedicine: Advancements in Herbal Products as Novel Drug Leads*. 2018.
95. Willis S, Sunkara R, Hester F, Shackelford L, Walker LT, Verghese M. Chemopreventive and Anti-Inflammatory Potential of Select Herbal Teas and Cinnamon in an &lt;i>In-Vitro&lt;/i> Cell Model. *Food Nutr Sci*. 2019;10(09).
96. Malik S, Saeed S, Saleem A, Khan MI, Khan A, Akhtar MF. Alternative treatment of polycystic ovary syndrome: pre-clinical and clinical basis for using plant-based drugs. Vol. 14, *Frontiers in Endocrinology*. 2023.
97. Onder A Yokakh. Medicinal Plants including Spices for the Treatment of Polycystic Ovary Syndrome (PCOS) with a Preclinical-Clinical Perspective and Phytotherapeutic Approaches in: *The Chemistry inside Spices & Herbs: Research and Development*. Chaurasia PK, Bharati SL, Singh S, editors. Vol. 3. Bentham Science Publishers; 2024. 1–45 p.
98. Wang C, Lv X, Liu W, Liu S, Sun Z. Uncovering the pharmacological mechanism of motherwort (*Leonurus japonicus* Houtt.) for treating menstrual disorders: A systems pharmacology approach. Vol. 89, *Computational Biology and Chemistry*. 2020.
99. Shang X, Pan H, Wang X, He H, Li M. *Leonurus japonicus* Houtt.: Ethnopharmacology, phytochemistry and pharmacology of an important traditional Chinese medicine. Vol. 152, *Journal of Ethnopharmacology*. 2014.
100. Yigit S, Yesilyurt I, Bitiktas S, Aksu Kilicle P, Duysak L, Yayla M, et al. Therapeutic effect of *Berberis vulgaris* fruit extract on histopathological changes and oxidative stress markers of ovarian ischemia and reperfusion injury in rats. *J King Saud Univ Sci*. 2024 Dec 1;36(11):103578.
101. Li X, Ullah I, Hou C, Liu Y, Xiao K. Network pharmacology and molecular docking study on the treatment of polycystic ovary syndrome with angelica sinensis- radix rehmanniae drug pair. *Medicine (United States)*. 2023;102(46).
102. Kim H, Choi E, Chung H, Joung Y, Shin D, Cho S. Effects of *Angelica Gigantis Radix* (AGR) on Polycystic

- Ovary induced by Estradiol Valerate in rats. *Planta Med.* 2011;77(12).
103. Zhu TW, Li XL. Berberine interacts with gut microbiota and its potential therapy for polycystic ovary syndrome. Vol. 50, *Clinical and Experimental Pharmacology and Physiology*. 2023.
  104. Wei W, Zhao H, Wang A, Sui M, Liang K, Deng H, et al. A clinical study on the short-term effect of berberine in comparison to metformin on the metabolic characteristics of women with polycystic ovary syndrome. *Eur J Endocrinol.* 2012;166(1).
  105. Bahmani F, Karamali M, Shakeri H, Asemi Z. The effects of folate supplementation on inflammatory factors and biomarkers of oxidative stress in overweight and obese women with polycystic ovary syndrome: A randomized, double-blind, placebo-controlled clinical trial. *Clin Endocrinol (Oxf).* 2014;81(4).
  106. Ciotta L, Stracquandano M, Pagano I, Carbonaro A, Palumbo M, Gulino F. Effects of Myo-Inositol supplementation on oocyte's quality in PCOS patients: A double blind trial. *Eur Rev Med Pharmacol Sci.* 2011;15(5).
  107. Genazzani AD. Inositol as putative integrative treatment for PCOS. Vol. 33, *Reproductive BioMedicine Online*. 2016.
  108. Unfer V, Carlomagno G, Dante G, Facchinetti F. Effects of myo-inositol in women with PCOS: A systematic review of randomized controlled trials. Vol. 28, *Gynecological Endocrinology*. 2012.
  109. Shi X ke, Peng T, Azimova B, Li X li, Li S shan, Cao D yi, et al. Luteolin and its analog luteolin-7-methylether from *Leonurus japonicus* Hoult suppress aromatase-mediated estrogen biosynthesis to alleviate polycystic ovary syndrome by the inhibition of tumor progression locus 2. *J Ethnopharmacol.* 2024 Sep 15;331:118279.
  110. Ong M, Cheng J, Jin X, Lao W, Johnson M, Tan Y, et al. Paeoniflorin extract reverses dexamethasone-induced testosterone over-secretion through downregulation of cytochrome P450 17A1 expression in primary murine theca cells. *J Ethnopharmacol.* 2019;229.
  111. Darabi P, Khazali H, Mehrabani Natanzi M. Therapeutic potentials of the natural plant flavonoid apigenin in polycystic ovary syndrome in rat model: via modulation of pro-inflammatory cytokines and antioxidant activity. *Gynecological Endocrinology.* 2020;36(7).
  112. Berk B, İlhan N, Susam S, Tedik F, Kaya Tektemur N. The ameliorating effects of apigenin and chrysin alone and in combination on polycystic ovary syndrome induced by dehydroepiandrosterone in rats. *Marmara Medical Journal.* 2024 May 31;37(2):198–207.
  113. Hong G, Wu H, Ma ST, Su Z. Catechins from oolong tea improve uterine defects by inhibiting STAT3 signaling in polycystic ovary syndrome mice. *Chinese Medicine (United Kingdom).* 2020;15(1).
  114. Sharafieh G, Salmanifarzaneh F, Gharbi N, Sarvestani FM, Rahmazad F, Razlighi MR, et al. Histological and molecular evaluation of *Mentha arvensis* extract on a polycystic ovary syndrome rat model. *J Bras Reprod Assist.* 2023;27(2).
  115. Van Barneveld RJ, Dunshea FR, Beddhu S, Filipowicz R, Chen XXWXX, Neilson JL, et al. Bone Health in Gynecologic Cancers-does FOSAVANCE Help? *American journal of clinical nutrition.* 2019;10(1).
  116. Alae S. Evaluation of the Effects of *Mentha Spicata* Extract on In-Vitro Maturation of Mouse Oocytes. *Journal of Advanced Medical Sciences and Applied Technologies.* 2016;2(2).
  117. Šarić-Kundalić B, Fialová S, Dobeš C, Ölzant S, Tekel'ová D, Grančai D, et al. Multivariate numerical taxonomy of *Mentha* species, hybrids, varieties and cultivars. *Sci Pharm.* 2009;77(4).
  118. Niksic H, Duric K, Omeragic E, Niksic H, Muratovic S, Becic F. Chemical characterization , antimicrobial and antioxidant properties of *Mentha spicata* L . ( Lamiaceae ) essential oil. *Bulletin of the Chemists and Tehcnologists of Bosnia and Herzegovina.* 2018;50(Lawrence 2007).
  119. Idm'hand E, Msanda F, Cherifi K. Ethnopharmacological review of medicinal plants used to manage diabetes in Morocco. *Clinical Phytoscience.* 2020;6(1).
  120. Mahboubi M. *Mentha spicata* L. essential oil, phytochemistry and its effectiveness in flatulence. Vol. 11, *Journal of Traditional and Complementary Medicine.* 2021.
  121. Bouyahya A, Abrini J, Et-Touys A, Bakri Y, Dakka N. Indigenous knowledge of the use of medicinal plants in the North-West of Morocco and their biological activities. *Eur J Integr Med.* 2017;13.
  122. Yousuf P. Analgesic, Anti-Inflammatory and Antipyretic Effect of *Mentha spicata* (Spearmint). *Br J Pharm Res.* 2013;3(4).
  123. Boukhebt H, Chaker AN, Belhadj H, Sahli F, Ramdhani M, Laouer H, et al. Chemical composition and antibacterial activity of *Mentha pulegium* L. and *Mentha spicata* L. essential oils. *Pharm Lett.* 2011;3(4).
  124. Zekri N, Elazzouzi H, Drioiche A, Zair T, El Belghiti MA. Antibacterial activity of essential oils and hydrosols extracted from some Moroccan *Mentha* species (L.). *Journal of Medical Pharmaceutical and Allied Sciences.* 2022;11(1).
  125. Abdul Jabbar AAS, Kathem SH. The protective effect of *Mentha spicata* ethanolic extract on irinotecan-induced mucositis in mice. *Iraqi Journal of Pharmaceutical Sciences.* 2019;28(1).
  126. Mehraban M, Jelodar G, Rahmanifar F. A combination of spearmint and flaxseed extract improved endocrine and histomorphology of ovary in experimental PCOS. *J Ovarian Res.* 2020;13(1).
  127. Grant P. Spearmint herbal tea has significant anti-androgen effects in polycystic ovarian syndrome. A randomized controlled trial. *Phytotherapy Research.* 2010;24(2).
  128. Bahl AS. Efficacy of Topical Application of Plant Extract Based Natural Oils in Management of Polycystic Ovarian Syndrome: a Two-Arm Parallel Group Randomised Double-Blind Placebo-Controlled Clinical Study. *Health Science Journal.* 2020;14(3):723.
  129. Özer Z. Investigation of Phenolic Compounds and Antioxidant Activity of *Mentha spicata* L. subsp. *spicata* and *M. longifolia* (L.) L. subsp. *typhoides* (Briq.) Harley Decoction and Infusion. *Journal of the Turkish Chemical Society Section A: Chemistry.* 2018 Jan 1;5(2):445–56.
  130. Nickavar B, Nickavar A. Compositional Analysis of Essential Oils from Two *Mentha* Species and in silico Study on their Major Volatile Constituents against Polycystic Ovary Syndrome. *Lett Drug Des Discov.* 2022;20(2).
  131. Çam M, Dinç Işıklı M, Yüksel E, Alaşalvar H, Başyigit B. Application of pressurized water extraction and spray drying techniques to produce soluble spearmint tea.

- Journal of Food Measurement and Characterization. 2018;12(3).
132. Ramanan S S, Arunachalam A, Singh R, Verdiya A. Tropical almond (*Terminalia catappa*): A holistic review. Vol. 11, Heliyon. Elsevier Ltd; 2025.
  133. Thomson LAJ, Evans B. Vol. ver.2.2, Species Profiles for Pacific Island Agroforestry. 2006. *Terminalia catappa* (tropical almond) Combretaceae (combretum family).
  134. : Flora of North America: North of Mexico. Volume 10: Magnoliophyta: Proteaceae to Elaeagnaceae . Q Rev Biol. 2022;97(4).
  135. Ladele B, Kpoviessi S, Ahissou H, Gbenou J, Kpadonou-Kpoviessi B, Mignolet E, et al. Chemical composition and nutritional properties of *Terminalia catappa* L. oil and kernels from Benin. Comptes Rendus Chimie. 2016;19(7).
  136. Anand AV, Divya N, Kotti PP. An updated review of *Terminalia catappa*. Vol. 9, Pharmacognosy Reviews. 2015.
  137. P V, V V, P B. Phytopharmacological significance of *terminalia catappa* l.: an updated review. Int J Res Ayurveda Pharm. 2016;7(2).
  138. Purohit A, Jain S, Nema P, Jain DK, Vishwakarma H, Jain PK. A Comprehensive Review on Tailoring an Herbal Approach for Treatment of Poly Cystic Ovarian Syndrome. Asian Journal of Dental and Health Sciences. 2022;2(1).
  139. Wal A, Wal P, Saraswat N, Wadhwa S. A Detailed Review on Herbal Treatments for Treatment of PCOS-Polycystic ovary syndrome (PCOS). Current Nutraceuticals. 2021;2(3).
  140. A Molecular Docking Study towards Finding Herbal Treatment against Polycystic Ovary Syndrome (PCOS). International Journal of Recent Technology and Engineering. 2020;8(2S12).
  141. Christian A, Ukhun ME. Nutritional potential of the nut of tropical almond (*Terminalia Catappia* L.). Pakistan Journal of Nutrition. 2006;5(4).
  142. Kasahara Y, Hemmi S. Medicinal herb index in Indonesia. 2nd ed. 1995. 1–453 p.
  143. Hung HD, Tien DD, Ngoan NT, Duong BT, Viet DQ, Dien PG, et al. Chemical constituents from the leaves of *terminalia catappa* l. (Combretaceae). Vietnam J Sci Technol. 2022;60(4).
  144. Oyeleye SI, Adebayo AA, Ogunsuyi OB, Dada FA, Oboh G. Phenolic profile and Enzyme Inhibitory activities of Almond (*Terminalia catappa*) leaf and Stem bark. Int J Food Prop. 2018;20.
  145. Abiodun OO, Rodríguez-Nogales A, Algieri F, Gomez-Caravaca AM, Segura-Carretero A, Utrilla MP, et al. Antiinflammatory and immunomodulatory activity of an ethanolic extract from the stem bark of *Terminalia catappa* L. (Combretaceae): In vitro and in vivo evidences. J Ethnopharmacol. 2016;192.
  146. Chen PS, Li JH, Liu TY, Lin TC. Folk medicine *Terminalia catappa* and its major tannin component, punicalagin, are effective against bleomycin-induced genotoxicity in Chinese hamster ovary cells. Cancer Lett. 2000;152(2).
  147. Chiang HM, Wen KC, Shih IC, Hu JC, Liao ST, Su TW. Inhibitory effects of *Terminalia catappa* on UVB-induced photodamage in fibroblast cell line. Evidence-based Complementary and Alternative Medicine. 2011;2011.
  148. Umarudeen AM, Magaji MG, Khan F, Abubakar A. Acute Antidepressant Activity Investigation of Selected African Medicinal Plants in Mice: A Preliminary Study. European Journal of Pharmaceutical Research. 2023;3(5).
  149. Danladi S, Muhammad MA, Yaro AH. Central nervous system depressant activity of ethanol leaf extract of *Terminalia braunii* (Engler) (Loranthaceae) growing on *Terminalia catappa* L. (Combretaceae). Bayero Journal of Pure and Applied Sciences. 2020;12(1).
  150. Chandrasekhar Y, Phani Kumar G, Navya K, Ramya EM, Anilakumar KR. Tannins from *Terminalia chebula* fruits attenuates GABA antagonist-induced anxiety-like behaviour via modulation of neurotransmitters. Journal of Pharmacy and Pharmacology. 2018;70(12).
  151. Braga FC, Serra CP, Viana Júnior NS, Oliveira AB, Côrtes SF, Lombardi JA. Angiotensin-converting enzyme inhibition by Brazilian plants. Fitoterapia. 2007;78(5).
  152. Ogbu PN, Famurewa AC, Ugbor CK, Ogbu IM, Aloke C, Obasi NA, et al. HPLC phytochemical profiling, antioxidant activity and in vitro evaluation of inhibitory effects of *Terminalia catappa* stem bark extract on enzymes linked to diabetes, hypertensive vasoconstriction and erectile dysfunction. Pharmacological Research - Natural Products. 2024 Sep 1;4:100064.
  153. Adefegha SA, Oboh G, Oyeleye SI, Ejakpovi I. Erectogenic, Antihypertensive, Antidiabetic, Anti-Oxidative Properties and Phenolic Compositions of Almond Fruit (*Terminalia catappa* L.) Parts (Hull and Drupe) – in vitro. J Food Biochem. 2017;41(2).
  154. Babalola OO, Iwaloye O, Ottu PO, Aturamu PO, Olawale F. Biological activities of African medicinal plants in the treatment of erectile dysfunction: a mechanistic perspective. Vol. 44, Hormone Molecular Biology and Clinical Investigation. 2023.
  155. Yadav S, Kapoor R, Mittal P, Ajmal G. *Terminalia catappa* Linn.: A treasury of pharmacological benefits. uttar pradesh journal of zoology. 2021;
  156. Japhet C.P. ., Luka C.D. ., Otitoju A.P. ., Miri P . Antilipidemic and Hepatorenal Effects of Aqueous Extracts of *Terminalia catappa* on Streptozotocin-induced Diabetic Rats. Asian Journal of Research in Medical and Pharmaceutical Sciences. 2024 May 18;13(2):72–8.
  157. Ambikadevi S, Mukundan U. Anti-obesity and anti-hyperlipidemic activity of *terminalia catappa* linn. in high-fat-diet induced obese rats. Asian Journal of Pharmaceutical Research and Development (An International Peer-Reviewed Journal of Pharmaceutical Research and Development [Internet]. 1(6):114–20. Available from: [www.ajprd.com](http://www.ajprd.com)[www.ajprd.com](http://www.ajprd.com)
  158. Ganer R, Kamble MA, Dhabarde DM, Ingole AR, Baheti JR. Evaluation of Analgesic and In-vitro Anti-Inflammatory potential of Fruit Flesh Extract of *Terminalia catappa* Linn. . Research Journal of Pharmacognosy and Phytochemistry. 2017;9(4).
  159. Daram P, Jitta SR, Shreedhara CS, Misra CS, Gourishetti K, Lobo R. Investigation of anti-inflammatory and anti-arthritic potentials of *Terminalia catappa* bark using in vitro assays and carrageenan-induced inflammation, complete Freund's adjuvant induced arthritis model in rats. South African Journal of Botany. 2021;141.
  160. Gao J, Dou H, Tang XH, Xu LZ, Fan YM, Zhao XN. Inhibitory effect of TCCE on CCL4-induced overexpression of IL-6 in acute liver injury. Acta Biochim Biophys Sin (Shanghai). 2004;36(11).

161. Abdul Vahab A, Harindran Jyoti. Hepatoprotective activity of bark extracts of Terminalia catappa linn in albino rats. *World J Pharm Pharm Sci.* 2016;5(6).
162. Tang X, Gao J, Wang Y, Fan YM, Xu LZ, Zhao XN, et al. Effective protection of Terminalia catappa L. leaves from damage induced by carbon tetrachloride in liver mitochondria. *Journal of Nutritional Biochemistry.* 2006;17(3).
163. Behl T, Kotwani A. Proposed mechanisms of Terminalia catappa in hyperglycaemia and associated diabetic complications. Vol. 69, *Journal of Pharmacy and Pharmacology.* 2017.
164. Hayaza S, Istiqomah S, Kuncoroningrat Susilo RJ, Inayatillah B, Ansori ANM, Winarni D, et al. Antidiabetic Activity of Ketapang (Terminalia catappa L.) Leaves Extract in Streptozotocin-Induced Diabetic Mice. *Indian Veterinary Journal.* 2019;96(12).
165. Gandhi PP, Venkatalakshmi P, Brindha P. Efficacy of Terminalia catappa L. Wood and Bark against Some Fungal Species [Internet]. Vol. 4, *Int.J.Curr.Microbiol. App.Sci.* 2015. Available from: <http://www.ijcmas.com>
166. Azrul LM, Adzemi MA, Effendy AWM, Imelda, Nurulaini R. Determination of Anthelmintic Potential in Terminalia catappa by Modified Selected In Vitro Bioassay. 2nd International Conference on Biotechnology and Food Science. 2011;7.
167. Mwangi WC, Waudu W, Shigwenya ME, Gichuki J. Phytochemical characterization, antimicrobial and antioxidant activities of Terminalia catappa methanol and aqueous extracts. *BMC Complement Med Ther.* 2024 Dec 1;24(1).
168. Taganna JC, Quanicco JP, Perono RMG, Amor EC, Rivera WL. Tannin-rich fraction from Terminalia catappa inhibits quorum sensing (QS) in Chromobacterium violaceum and the QS-controlled biofilm maturation and LasA staphylolytic activity in Pseudomonas aeruginosa. *J Ethnopharmacol.* 2011;134(3).
169. Akharaiyi FC, Ilori RM, Adesida JA. International Journal of Pharmaceutical and Biomedical Research Antibacterial effect of Terminalia catappa on some selected pathogenic bacteria. 2011.
170. Ibegbulem CO, Essien. Biochemical effects of drinking Terminalia catappa Linn. decoction in Wistar rats. *African Journal of Biochemistry Research.* 2011;5(8).
171. Aimola IA, Inuwa HM, Nok AJ, Mamman AI. Induction of foetal haemoglobin synthesis in erythroid progenitor stem cells: Mediated by water-soluble components of Terminalia catappa. *Cell Biochem Funct.* 2014;32(4).
172. Nugroho RA, Utami D, Aryani R, Nur FM, Sari YP, Manurung H. In vivo wound healing activity of ethanolic extract of Terminalia catappa L. leaves in mice (Mus musculus). In: *Journal of Physics: Conference Series.* 2019.
173. Ariani L, Desmiaty Y. Antioxidant and anti-elastase activity of Tampui (Baccaurea macrocarpa) and Ketapong (Terminalia badamia) barks and leaves. 2024; Available from: <https://www.researchgate.net/publication/386110587>
174. Venkatalakshmi P, Brindha P, Induja K. In -vitro anti oxidant and antitumor studies on terminalia catappa bark. *Int J Pharm Pharm Sci.* 2014;6(SUPPL 1).
175. Ko TF, Weng YM, Lin S Bin, Chiou RYY. Antimutagenicity of supercritical CO<sub>2</sub> extracts of Terminalia catappa leaves and cytotoxicity of the extracts to human hepatoma cells. *J Agric Food Chem.* 2003;51(12).
176. Chu SC, Yang SF, Liu SJ, Kuo WH, Chang YZ, Hsieh YS. In vitro and in vivo antimetastatic effects of Terminalia catappa L. leaves on lung cancer cells. *Food and Chemical Toxicology.* 2007;45(7).
177. Ayeni EA, Gong Y, Yuan H, Hu Y, Bai X, Liao X. Medicinal Plants for Anti-neurodegenerative diseases in West Africa. Vol. 285, *Journal of Ethnopharmacology.* 2022.
178. Venkatalakshmi P, Brindha P, Vadivel V. In vitro Antioxidant and Anti-inflammatory Studies on Bark, Wood and Fruits of Terminalia catappa L. *International Journal of Phytomedicine.* 2015;7(3).
179. Sivaranjani C, Venkatalakshmi P, Brindha P. In Vitro Anti Inflammatory and Antioxidant Activities on Fruits of Terminalia catappa L. . *Res J Pharm Technol.* 2015;8(10).
180. Olatidoye OP, Sobowale SS, Akinlotan J V, Olorode OO. chemical composition and physicochemical characteristics of tropical almond nuts (terminalia catappa l) cultivated in south west nigeria. *M e d i c a l a n d A p p l i e d B i o s c i e n c e s.* 2011;2(March).
181. Gutiérrez-Pineda KM, Herrera M. Preliminary phytochemical characterization of the ethanolic extracts of leaf, green and ripe fruit of terminalia catappa l. (almendro) in panama. *Granja.* 2022;36(2).
182. Suryavanshi A, Saxena AM. Phytochemicals: Extraction and Preliminary Investigation of Bioactive Compounds from Rumex Vesicarius and Terminalia Catappa Plant. *Asian Journal of Engineering and Applied Technology.* 2019;8(3).
183. Danladi S, Sule MI, Muhammad MA, Yaro AH. Isolation and Characterization of Some Flavonoids from the Leaves of Globimetula braunii (Loranthaceae) Growing on Terminalia catappa L. (Combretaceae). *Tropical Journal of Natural Product Research.* 2022;5(12).
184. Lokesh Ravi, Divya Jindam, Suganya KUMARESAN, Venkatesh Selvaraj, Jayarama Reddy. Anti-methicillin resistant staphylococcus aureus potential of phytochemicals in terminalia catappa and their proposed in silico mechanism of action. *Asian Journal of Pharmaceutical and Clinical Research.* 2019;
185. Ram J, Moteriya P, Chanda S, Chanda Phytochemical S. Phytochemical screening and reported biological activities of some medicinal plants of Gujarat region. ~ 192 ~ *Journal of Pharmacognosy and Phytochemistry.* 2015;4(2).
186. Muthulakshmi L, Suganya K, Murugan M, Annaraj J, Duraipandiyan V, Al Farraj DA, et al. Antibiofilm efficacy of novel biogenic silver nanoparticles from Terminalia catappa against food-borne Listeria monocytogenes ATCC 15,313 and mechanisms investigation in-vivo and in-vitro. *J King Saud Univ Sci.* 2022;34(5).
187. Tata B, Mimouni NEH, Barbotin AL, Malone SA, Loyens A, Pigny P, et al. Elevated prenatal anti-Müllerian hormone reprograms the fetus and induces polycystic ovary syndrome in adulthood. *Nat Med.* 2018;24(6).
188. Sanchez-Garrido MA, Tena-Sempere M. Metabolic dysfunction in polycystic ovary syndrome: Pathogenic role of androgen excess and potential therapeutic strategies. Vol. 35, *Molecular Metabolism.* 2020.

189. Atanassova N, Koev Y. Hydrohysteroid Dehydrogenases – Biological Role and Clinical Importance – Review. In: Dehydrogenases. 2012.
190. Chung HJ, Noh Y, Kim MS, Jang A, Lee CE, Myung SC. Steroidogenic effects of Taraxacum officinale extract on the levels of steroidogenic enzymes in mouse Leydig cells. *Anim Cells Syst* (Seoul). 2018;22(6).
191. Payne AH, Youngblood GL. Regulation of expression of steroidogenic enzymes in Leydig cells. In: *Biology of Reproduction*. 1995.
192. Arakane F, Kallen CB, Watari H, Foster JA, Sepuri NBV, Pain D, et al. The mechanism of action of steroidogenic acute regulatory protein (StAR): StAR acts on the outside of mitochondria to stimulate steroidogenesis. *Journal of Biological Chemistry*. 1998;273(26).
193. Moran LJ, Brown WJ, McNaughton S, Joham AE, Teede HJ. Weight management practices associated with polycystic ovary syndrome and their relationships with diet and physical activity. *Obes Res Clin Pract*. 2019;13(1).
194. Pirota S, Lim SS, Grassi A, Couch LM, Jeanes YM, Joham AJ, et al. Relationships between self-management strategies and physical activity and diet quality in women with polycystic ovary syndrome. *Patient Educ Couns*. 2022;105(1).
195. Tatone C, Di Emidio G, Placidi M, Rossi G, Ruggieri S, Taccaliti C, et al. AGEs-related dysfunctions in PCOS: Evidence from animal and clinical research. Vol. 251, *Journal of Endocrinology*. 2021.
196. Merhi Z, Kandaraki EA, Diamanti-Kandarakis E. Implications and Future Perspectives of AGEs in PCOS Pathophysiology. Vol. 30, *Trends in Endocrinology and Metabolism*. 2019.
197. Marianian AY, Kuzmin MY, Markova DP. Ego-Identity of Women in the Reproductive Age with Pcos and Their Socio-Demographic Determinants. *J Pharm Res Int*. 2020;
198. Scarfò G, Daniele S, Fusi J, Gesi M, Martini C, Franzoni F, et al. Metabolic and Molecular Mechanisms of Diet and Physical Exercise in the Management of Polycystic Ovarian Syndrome. Vol. 10, *Biomedicines*. 2022.
199. IRCT20180714040462N1, ISRCTN82088636, Brodner DC, Corsino P, Harvey A, Souètre E, et al. Effect of blue light from electronic devices on melatonin and sleep/wake rhythms in high school children. *Sleep*. 2020;40(1).
200. Tao Y, Liu B, Chen Y, Hu Y, Zhu R, Ye D, et al. Genetically predicted cigarette smoking in relation to risk of polycystic ovary syndrome. *Clin Epidemiol*. 2021;13.
201. Shele G, Genkil J, Speelman D. A systematic review of the effects of exercise on hormones in women with polycystic ovary syndrome. Vol. 5, *Journal of Functional Morphology and Kinesiology*. 2020.
202. Butt MS, Saleem J, Zakar R, Aiman S, Khan MZ, Fischer F. Benefits of physical activity on reproductive health functions among polycystic ovarian syndrome women: a systematic review. *BMC Public Health*. 2023;23(1).
203. Xirofotos D, Trakakis E, Peppas M, Chrelias C, Panagopoulos P, Christodoulaki C, et al. The amount and duration of smoking is associated with aggravation of hormone and biochemical profile in women with PCOS. *Gynecological Endocrinology*. 2016;32(2).
204. Mimouni NEH, Paiva I, Barbotin AL, Timzoura FE, Plassard D, Le Gras S, et al. Polycystic ovary syndrome is transmitted via a transgenerational epigenetic process. *Cell Metab*. 2021;33(3).
205. Attea BMR, El-Kak AEA, Lucchesi PA, Delafontane P. Antioxidant activity of folic acid: From mechanism of action to clinical application. *The FASEB Journal*. 2009;23(S1).
206. Miraglia N, Dehay E. Folate Supplementation in Fertility and Pregnancy: The Advantages of (6S)5-Methyltetrahydrofolate. *Altern Ther Health Med*. 2022;28(4).
207. Rahimi S, Martel J, Karahan G, Angle C, Behan NA, Chan D, et al. Moderate maternal folic acid supplementation ameliorates adverse embryonic and epigenetic outcomes associated with assisted reproduction in a mouse model. *Human Reproduction*. 2019;34(5).
208. Gliszczynska-Świgło A. Folates as antioxidants. *Food Chem*. 2007;101(4).
209. Ebisch IMW, Thomas CMG, Peters WHM, Braat DDM, Steegers-Theunissen RPM. The importance of folate, zinc and antioxidants in the pathogenesis and prevention of subfertility. Vol. 13, *Human Reproduction Update*. 2007.
210. Racek J, Rusňáková H, Trefil L, Siala KK. The influence of folate and antioxidants on homocysteine levels and oxidative stress in patients with hyperlipidemia and hyperhomocysteinemia. *Physiol Res*. 2005;54(1).
211. Jafari A, Gholizadeh E, Sadrmanesh O, Tajpour S, Yarizadeh H, Zamani B, et al. The effect of folic acid supplementation on body weight and body mass index: A systematic review and meta-analysis of randomized controlled trials. *Clin Nutr ESPEN*. 2023;53.
212. Sobral AF, Cunha A, Silva V, Gil-Martins E, Silva R, Barbosa DJ. Unveiling the Therapeutic Potential of Folate-Dependent One-Carbon Metabolism in Cancer and Neurodegeneration. *Int J Mol Sci* [Internet]. 2024 Aug 28 [cited 2025 Jan 24];25(17):9339. Available from: <https://www.mdpi.com/1422-0067/25/17/9339>
213. Durga J, Bots ML, Schouten EG, Grobbee DE, Kok FJ, Verhoef P. Effect of 3 y of folic acid supplementation on the progression of carotid intima-media thickness and carotid arterial stiffness in older adults. *American Journal of Clinical Nutrition*. 2011;93(5).
214. Antoniadou C. Homocysteine and coronary atherosclerosis: from folate fortification to recent clinical trials. *Free Radic Biol Med*. 2013;65.
215. Molloy AM, Scott JM. Folates and prevention of disease. *Public Health Nutr*. 2001;4(2b).
216. Palomba S, Falbo A, Giallauria F, Russo T, Tolino A, Zullo F, et al. Effects of metformin with or without supplementation with folate on homocysteine levels and vascular endothelium of women with polycystic ovary syndrome. *Diabetes Care*. 2010;33(2).
217. Paquette AF, Carbone BE, Vogel S, Israel E, Maria SD, Patil NP, et al. The human milk component myo-inositol promotes neuronal connectivity. *Proc Natl Acad Sci U S A*. 2023;120(30).
218. Chhetri DR. Myo-inositol and its derivatives: Their emerging role in the treatment of human diseases. Vol. 10, *Frontiers in Pharmacology*. 2019.
219. Packer L, Witt EH, Tritschler HJ. Alpha-lipoic acid as a biological antioxidant. *Free Radic Biol Med*. 1995 Aug 1;19(2):227–50.

220. Wojcik M, Mac-Marcjanek K, Wozniak L. Physiological and Pathophysiological Functions of SIRT1. *Mini-Reviews in Medicinal Chemistry*. 2012;9(3).
221. Chatree S, Thongmaen N, Tantivejkul K, Sitticharoon C, Vucenik I. Role of inositols and inositol phosphates in energy metabolism. Vol. 25, *Molecules*. 2020.
222. Watkins OC, Yong HEJ, Sharma N, Chan SY. A review of the role of inositols in conditions of insulin dysregulation and in uncomplicated and pathological pregnancy. Vol. 62, *Critical Reviews in Food Science and Nutrition*. 2022.
223. Benvenga S, Nordio M, Laganà AS, Unfer V. The Role of Inositol in Thyroid Physiology and in Subclinical Hypothyroidism Management. Vol. 12, *Frontiers in Endocrinology*. 2021.
224. Bizzarri M, Monti N, Piombarolo A, Angeloni A, Verna R. Myo-Inositol and D-Chiro-Inositol as Modulators of Ovary Steroidogenesis: A Narrative Review. Vol. 15, *Nutrients*. 2023.
225. Siracusa L, Napoli E, Ruberto G. Novel Chemical and Biological Insights of Inositol Derivatives in Mediterranean Plants. Vol. 27, *Molecules*. 2022.
226. KIANI AK, PAOLACCI S, CALOGERO AE, CANNARELLA R, DI RENZO GC, GERLI S, et al. From Myo-inositol to D-chiro-inositol molecular pathways. *Eur Rev Med Pharmacol Sci*. 2021;25(5).
227. Park S, Karunakaran U, Jeoung N, Jeon JH, Lee IK. Physiological Effect and Therapeutic Application of Alpha Lipoic Acid. *Curr Med Chem*. 2014;21(32).
228. Viana MDM, Lauria PSS, Lima AA de, Opretzka LCF, Marcelino HR, Villarreal CF. Alpha-Lipoic Acid as an Antioxidant Strategy for Managing Neuropathic Pain. Vol. 11, *Antioxidants*. 2022.
229. Brizzi A, Maramai S, Aiello F, Baratto MC, Corelli F, Mugnaini C, et al. Lipoic/Capsaicin-Related Amides: Synthesis and Biological Characterization of New TRPV1 Agonists Endowed with Protective Properties against Oxidative Stress. *Int J Mol Sci*. 2022;23(21).
230. Kamenov Z, Gateva A. Inositols in pcos. Vol. 25, *Molecules*. 2020.
231. Greff D, Juhász AE, Vánca S, Váradi A, Sipos Z, Szinte J, et al. Inositol is an effective and safe treatment in polycystic ovary syndrome: a systematic review and meta-analysis of randomized controlled trials. Vol. 21, *Reproductive Biology and Endocrinology*. 2023.
232. Capece U, Moffa S, Improta I, Di Giuseppe G, Nista EC, Cefalo CMA, et al. Alpha-Lipoic Acid and Glucose Metabolism: A Comprehensive Update on Biochemical and Therapeutic Features. Vol. 15, *Nutrients*. 2023.
233. Ko CY, Wu CH, Huang WJ, Lo YM, Lin SX, Wu JSB, et al. Alleviative effects of  $\alpha$ -lipoic acid on muscle atrophy via the modulation of TNF- $\alpha$ /JNK and PI3K/AKT pathways in high-fat diet and streptozotocin-induced type 2 diabetic rats. *Food Sci Nutr*. 2023;11(4).
234. Tóth F, Cseh EK, Vécsei L. Natural molecules and neuroprotection: Kynurenic acid, pantethine and  $\alpha$ -lipoic acid. Vol. 22, *International Journal of Molecular Sciences*. 2021.
235. Genazzani AD, Shefer K, Della Casa D, Prati A, Napolitano A, Manzo A, et al. Modulatory effects of alpha-lipoic acid (ALA) administration on insulin sensitivity in obese PCOS patients. *J Endocrinol Invest*. 2018;41(5).
236. Cappelli V, Di Sabatino A, Musacchio MC, De Leo V. [Evaluation of a new association between insulin-sensitizers and  $\alpha$ -lipoic acid in obese women affected by PCOS]. *Minerva Ginecol*. 2013;65(4).
237. Cirillo F, Catellani C, Lazzeroni P, Sartori C, Tridenti G, Vezzani C, et al. HMGB1 is increased in adolescents with polycystic ovary syndrome (PCOS) and decreases after treatment with myo-inositol (MYO) in combination with alpha-lipoic acid (ALA). *Gynecological Endocrinology*. 2020;36(7).
238. Artini PG, Obino MER, Micelli E, Malacarne E, Vacca C, Papini F, et al. Effect of d-chiro-inositol and alpha-lipoic acid combination on COH outcomes in overweight/obese PCOS women. *Gynecological Endocrinology*. 2020;36(9).
239. Cianci A, Panella M, Fichera M, Falduzzi C, Bartolo M, Caruso S. D-chiro-Inositol and alpha lipoic acid treatment of metabolic and menses disorders in women with PCOS. *Gynecological Endocrinology*. 2015;31(6).
240. Rago R, Marcucci I, Leto G, Caponecchia L, Salacone P, Bonanni P, et al. Effect of myo-inositol and alpha-lipoic acid on oocyte quality in polycystic ovary syndrome non-obese women undergoing in vitro fertilization: A pilot study. *J Biol Regul Homeost Agents*. 2015;29(4).
241. Chang HC, Guarente L. SIRT1 and other sirtuins in metabolism. Vol. 25, *Trends in Endocrinology and Metabolism*. 2014. p. 138–45.
242. Ng F, Wijaya L, Tang BL. SIRT1 in the brain—connections with aging-associated disorders and lifespan. Vol. 9, *Frontiers in Cellular Neuroscience*. 2015.
243. Revollo JR, Li X. The ways and means that fine tune Sirt1 activity. Vol. 38, *Trends in Biochemical Sciences*. 2013.
244. Carafa V, Rotili D, Forgione M, Cuomo F, Serretiello E, Hailu GS, et al. Sirtuin functions and modulation: from chemistry to the clinic. Vol. 8, *Clinical Epigenetics*. 2016.
245. Yamamoto H, Schoonjans K, Auwerx J. Sirtuin functions in health and disease. Vol. 21, *Molecular Endocrinology*. 2007.
246. Curry AM, White DS, Donu D, Cen Y. Human Sirtuin Regulators: The “Success” Stories. Vol. 12, *Frontiers in Physiology*. 2021.
247. Khaidizar FD, Nakahata Y, Kume A, Sumizawa K, Kohno K, Matsui T, et al. Nicotinamide phosphoribosyltransferase delays cellular senescence by upregulating SIRT1 activity and antioxidant gene expression in mouse cells. *Genes to Cells*. 2017;22(12).
248. Fusi J, Bianchi S, Daniele S, Pellegrini S, Martini C, Galetta F, et al. An in vitro comparative study of the antioxidant activity and SIRT1 modulation of natural compounds. *Biomedicine and Pharmacotherapy*. 2018;101.
249. Terriente-Palacios C, Rubiño S, Hortós M, Peteiro C, Castellari M. Taurine, homotaurine, GABA and hydrophobic amino acids content influences “in vitro” antioxidant and SIRT1 modulation activities of enzymatic protein hydrolysates from algae. *Sci Rep*. 2022;12(1).
250. Di Emidio G, Placidi M, Rea F, Rossi G, Falone S, Cristiano L, et al. Methylglyoxal-dependent glycative stress and deregulation of SIRT1 functional network in the ovary of PCOS mice. *Cells*. 2020;9(1).
251. Pasquariello R, Verdile N, Brevini TAL, Gandolfi F, Boiti C, Zerani M, et al. The role of resveratrol in mammalian reproduction. Vol. 25, *Molecules*. 2020.
252. Wu YX, Yang XY, Han BS, Hu YY, An T, Lv BH, et al. Naringenin regulates gut microbiota and SIRT1/PGC-1 $\alpha$  signaling pathway in rats with letrozole-induced

- polycystic ovary syndrome. *Biomedicine and Pharmacotherapy*. 2022;153.
253. Fujita Y, Yamashita T. Sirtuins in neuroendocrine regulation and neurological diseases. Vol. 12, *Frontiers in Neuroscience*. 2018.
254. Jiao F, Gong Z. The Beneficial Roles of SIRT1 in Neuroinflammation-Related Diseases. Vol. 2020, *Oxidative Medicine and Cellular Longevity*. 2020.
255. Nakahata Y, Kaluzova M, Grimaldi B, Sahar S, Hirayama J, Chen D, et al. The NAD<sup>+</sup>-Dependent Deacetylase SIRT1 Modulates CLOCK-Mediated Chromatin Remodeling and Circadian Control. *Cell*. 2008;134(2).
256. Nayagam VM, Wang X, Yong CT, Poulsen A, Kee CG, Ng T, et al. SIRT1 modulating compounds from high-throughput screening as anti-inflammatory and insulin-sensitizing agents. *J Biomol Screen*. 2006;11(8).
257. Yang Y, Liu Y, Wang Y, Chao Y, Zhang J, Jia Y, et al. Regulation of SIRT1 and Its Roles in Inflammation. Vol. 13, *Frontiers in Immunology*. 2022.
258. Lu Z, Chen C, Gao Y, Li Y, Zhao X, Zhang H, et al. Screening target genes for the treatment of PCOS via analysis of single-cell sequencing data. *Ann Med*. 2022;54(1).
259. Zhao J, Tan Y, Feng Z, Zhou Y, Wang F, Zhou G, et al. Catalpol attenuates polycystic ovarian syndrome by regulating sirtuin 1 mediated NF- $\kappa$ B signaling pathway. *Reprod Biol*. 2022;22(3).
260. NCT04867252. Effects of Combined Resveratrol and Myo-inositol on Altered Metabolic, Endocrine Parameters and Perceived Stress in Patients With Polycystic Ovarian Syndrome. <https://clinicaltrials.gov/show/NCT04867252>. 2021;
261. Tatone C, di Emidio G, Barbonetti A, Carta G, Luciano AM, Falone S, et al. Sirtuins in gamete biology and reproductive physiology: Emerging roles and therapeutic potential in female and male infertility. *Hum Reprod Update*. 2018;24(3).
262. Wu M, Zhang J, Gu R, Dai F, Yang D, Zheng Y, et al. The role of Sirtuin 1 in the pathophysiology of polycystic ovary syndrome. Vol. 27, *European Journal of Medical Research*. 2022.
263. Lodge JK, Youn HD, Handelman GJ, Konishi T, Matsugo S, Mathur V V., et al. Natural sources of lipoic acid: Determination of lipoyllysine released from protease-digested tissues by high performance liquid chromatography incorporating electrochemical detection. *Journal of Applied Nutrition*. 1997;49(1-2).
264. Xiao R, Wang X, Jiang L, Tang H. Research and Application of Lipoic Acid in Plants. In: *IOP Conference Series: Earth and Environmental Science*. 2018.
265. Huo P, Li M, Le J, Zhu C, Yao J, Zhang S. Resveratrol improves follicular development of PCOS rats via regulating glycolysis pathway and targeting SIRT1. *Syst Biol Reprod Med*. 2023;69(2).
266. Wiciński M, Erdmann J, Nowacka A, Kuźmiński O, Michalak K, Janowski K, et al. Natural Phytochemicals as SIRT Activators—Focus on Potential Biochemical Mechanisms. Vol. 15, *Nutrients*. 2023.
267. Mahendran G, Verma SK, Rahman LU. The traditional uses, phytochemistry and pharmacology of spearmint (*Mentha spicata* L.): A review. Vol. 278, *Journal of Ethnopharmacology*. 2021.
268. Vahed Mohammad A, Author C, Vahed Mohammad Address -Abdul. Effect Of Terminalia Catappa On Hypoxia And Chronic Cold Resistance Stress In Experimental Animals.
269. Fatiha B, Khodir M, Farid D, Tiziri R, Karima B, Sonia O, et al. Pharmacognosy Communications Optimisation Of Solvent Extraction Of Antioxidants (Phenolic Compounds) From Algerian Mint ( *Mentha spicata* L. ). 2. Available from: [www.phcogcommn.org](http://www.phcogcommn.org)
270. Ethirajulu A, Alkasabera A, Onyali CB, Anim-Koranteng C, Shah HE, Bhawnani N, et al. Insulin Resistance, Hyperandrogenism, and Its Associated Symptoms Are the Precipitating Factors for Depression in Women With Polycystic Ovarian Syndrome. *Cureus*. 2021;
271. Alur-Gupta S, Dokras A. Considerations in the Treatment of Depression and Anxiety in Women with PCOS. *Semin Reprod Med*. 2023;41(1-2).
272. Dybciak P, Humeniuk E, Raczkiewicz D, Krakowiak J, Wdowiak A, Bojar I. Anxiety and Depression in Women with Polycystic Ovary Syndrome. *Medicina (Lithuania)*. 2022;58(7).
273. Cooney LG, Milman LW, Hantsoo L, Kornfield S, Sammel MD, Allison KC, et al. Cognitive-behavioral therapy improves weight loss and quality of life in women with polycystic ovary syndrome: a pilot randomized clinical trial. *Fertil Steril*. 2018 Jul 1;110(1):161-171.e1.
274. Jiskoot G, Timman R, Beerthuisen A, Dietz de Loos A, Busschbach J, Laven J. Weight Reduction Through a Cognitive Behavioral Therapy Lifestyle Intervention in PCOS: The Primary Outcome of a Randomized Controlled Trial. *Obesity*. 2020 Nov 1;28(11):2134-41.
275. Jiskoot G, Benneheij SH, Beerthuisen A, De Niet JE, De Klerk C, Timman R, et al. A three-component cognitive behavioural lifestyle program for preconceptional weight-loss in women with polycystic ovary syndrome (PCOS): A protocol for a randomized controlled trial. *Reprod Health*. 2017 Mar 6;14(1).

\*\*\*\*\*